

Inferring Latent Functions with Gaussian Processes in Differential Equations

Neil Lawrence

School of Computer Science
University of Manchester

Joint work with **Magnus Rattray** and **Guido Sanguinetti**

9th December 2006

Outline

- 1 Application
 - Transcription Factor Concentrations
- 2 Linear Response Model
 - Exact Inference in Linear System
 - Toy Problem
 - Biological Problem
- 3 Non-linear Response Model
 - Riemann Quadrature
 - Linear Response with MLP Kernel
 - Non-linear Responses

Advert!

PUMA Project

- Work is part of PUMA project for Microarray analysis
 - <http://bioinf.man.ac.uk/resources/puma>
- Will be appointing a new Post-doc in the new year!

Online Resources

All source code and slides are available online

- This talk available from home page (see talks link on side).
- Scripts available in the 'gpsim' toolbox
 - `http://www.dcs.shef.ac.uk/~neil/gpsim/`.
- MATLAB commands used for examples given in typewriter font.

Gaussian Processes

Inference about functions

- Differential equation models of systems often have functional unknowns.
 - We will focus on a protein concentration example.
- Gaussian processes (GPs) are probabilistic models for functions. O'Hagan [1978, 1992], Rasmussen and Williams [2006]
- GPs provide a framework for performing inference about these functions in the presence of uncertainty.

Application Overview

Transcription Factor Inference

- Model the dynamics of gene transcription.
- Infer transcription factor concentration (TFC).
- Infer parameters of the transcription model (decay rates, sensitivities *etc.*)
- Infer these quantities using a set of known target genes.

Methodology

Latent function

- Treat TFC as a *latent function* in a differential equation model.
- Assume a Gaussian process (GP) prior distribution for the latent function.
- Derive GP covariance jointly for genes and transcription factor.
- Maximise likelihood with respect to parameters (mostly physically meaningful).

Overview

Application Background

- Understanding of cellular processes is improving through microarrays, chromatin immunoprecipitation *etc.*.
- Quantitative description of regulatory mechanisms requires:
 - transcription factor (TF) concentrations,
 - gene-specific constants such as the baseline expression, mRNA decay rate and sensitivity to TF concentrations (TFCs).

Overview

Application Background II

- These quantities are hard to *measure directly*.
- We show how the
- They can be *inferred* using a systems biology model and Gaussian processes (GPs).
- Our work provides a extensible, principled framework for attacking this problem.

Gaussian Processes

GP Advantages

- GPs allow for inference of continuous profiles, accounting naturally for temporal structure.
 - GPs avoid cumbersome interpolation to estimate mRNA production rates.
 - GPs deal consistently with the uncertainty inherent in the measurements.
 - GPs outstrip MCMC for computational efficiency.
- GPs have previously been proposed for solving differential equations [Graepel, 2003] and dynamical systems [Murray-Smith and Pearlmutter].

Linear Response Model

p53 Inference [Barenco et al., 2006]

- Data consists of T measurements of mRNA expression level for N different genes.
- We relate gene expression, $x_j(t)$, to TFC, $f(t)$, by

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t). \quad (1)$$

B_j basal transcription rate of gene j ,

S_j is sensitivity of gene j

D_j is the decay rate of the mRNA.

- Dependence of mRNA transcription rate on TF is linear.

Linear Response Solution

Solve for TFC

- The equation given in (1) can be solved to recover

$$x_j(t) = \frac{B_j}{D_j} + S_j \exp(-D_j t) \int_0^t f(u) \exp(D_j u) du. \quad (2)$$

- If we model $f(t)$ as a GP then as (2) only involves linear operations $x_j(t)$ is also a GP.
- If we use the RBF covariance everything is analytic.

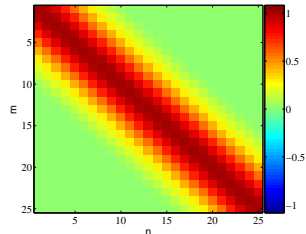
Covariance Functions

Visualisation of RBF Covariance

RBF Kernel Function

$$k(t, t') = \exp\left(-\frac{(t - t')^2}{l^2}\right)$$

- Covariance matrix is built using the time *inputs* to the function.
- For the example above it was based on Euclidean distance.
- The covariance function is also known as a kernel.



Covariance Samples

demCovFuncSample (oxford toolbox)

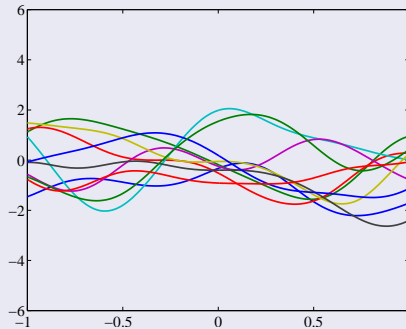


Figure: RBF kernel with $l = 0.25$, $\alpha = 1$

Covariance Samples

demCovFuncSample (oxford toolbox)

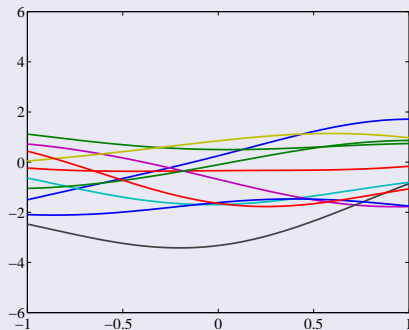


Figure: RBF kernel with $l = 1$, $\alpha = 1$

Computation of Joint Covariance

Covariance Function Computation

- We rewrite equation (2) as

$$x_j(t) = \frac{B_j}{D_j} + L_j[f](t)$$

where

$$L_j[f](t) = S_j \exp(-D_j t) \int_0^t f(u) \exp(D_j u) du \quad (3)$$

is a linear operator.

Induced Covariance

Gene's Covariance

- The new covariance function is then given by

$$\text{cov} (L_j [f] (t), L_k [f] (t')) = L_j \otimes L_k [k_{ff}] (t, t').$$

more explicitly

$$\begin{aligned} k_{x_j x_k} (t, t') &= S_j S_k \exp (-D_j t - D_k t') \int_0^t \exp (D_j u) \\ &\quad \times \int_0^{t'} \exp (D_k u') k_{ff} (u, u') du' du. \end{aligned}$$

- With RBF covariance these integrals are tractable.

Covariance Result

Covariance Result

$$k_{x_j x_k}(t, t') = S_j S_k \frac{\sqrt{\pi}}{2} [h_{kj}(t', t) + h_{jk}(t, t')]$$

where

$$h_{kj}(t', t) = \frac{\exp(\gamma_k)^2}{D_j + D_k} \\ \times \left\{ \exp[-D_k(t' - t)] \left[\operatorname{erf}\left(\frac{t' - t}{l} - \gamma_k\right) + \operatorname{erf}\left(\frac{t}{l} + \gamma_k\right) \right] \right. \\ \left. - \exp[-(D_k t' + D_j)] \left[\operatorname{erf}\left(\frac{t'}{l} - \gamma_k\right) + \operatorname{erf}(\gamma_k) \right] \right\}.$$

Here $\gamma_k = \frac{D_k l}{2}$.

Cross Covariance

Correlation of $x_j(t)$ and $f(t')$

- Need the “cross-covariance” terms between $x_j(t)$ and $f(t')$, which is obtained as

$$k_{x_j f}(t, t') = S_j \exp(-D_j t) \int_0^t \exp(D_j u) k_{ff}(u, t') du. \quad (4)$$

- For RBF we have

$$k_{x_j f}(t', t) = \frac{\sqrt{\pi} S_j e^{2\gamma_j}}{2} \exp[-D_j(t' - t)] \left[\operatorname{erf}\left(\frac{t' - t}{l} - \gamma_j\right) + \operatorname{erf}\left(\frac{t}{l} + \gamma_j\right) \right].$$

Posterior for f

Prediction for TFC

- Standard Gaussian process regression techniques [see e.g. Rasmussen and Williams, 2006] yield

$$\langle f \rangle_{\text{post}} = K_{f\mathbf{x}} K_{\mathbf{xx}}^{-1} \mathbf{x}$$
$$K_{ff}^{\text{post}} = K_{ff} - K_{f\mathbf{x}} K_{\mathbf{xx}}^{-1} K_{\mathbf{x}f}$$

- Model parameters B_j , D_j and S_j estimated by type II maximum likelihood,

$$\log p(\mathbf{x}) = N(\mathbf{x} | \mathbf{0}, K_{\mathbf{xx}})$$

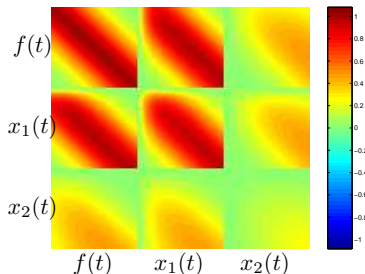
Covariance for Transcription Model

RBF Kernel function for $f(t)$

$$x_i(t) = \frac{B_i}{D_i} + S_i \exp(-D_i t) \int_0^t f(u) \exp(D_i u) du.$$

- Joint distribution for $x_1(t)$, $x_2(t)$ and $f(t)$.
- Here:

D_1	S_1	D_2	S_2
5	5	0.5	0.5



Joint Sampling of $x(t)$ and $f(t)$

gpsimTest

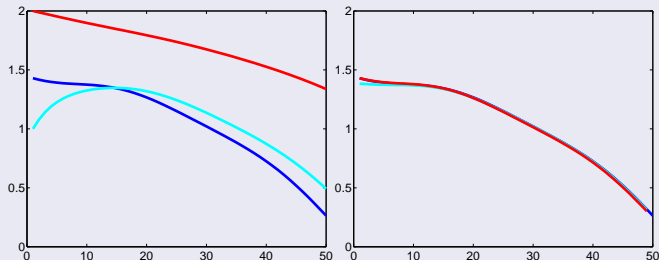


Figure: *Left:* joint samples from the transcription covariance, *blue:* $f(t)$, *cyan:* $x_1(t)$ and *red:* $x_2(t)$. *Right:* numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

Joint Sampling of $x(t)$ and $f(t)$

gpsimTest

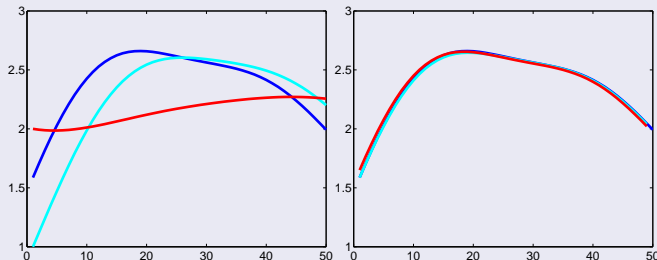


Figure: *Left:* joint samples from the transcription covariance, *blue:* $f(t)$, *cyan:* $x_1(t)$ and *red:* $x_2(t)$. *Right:* numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

Joint Sampling of $x(t)$ and $f(t)$

gpsimTest

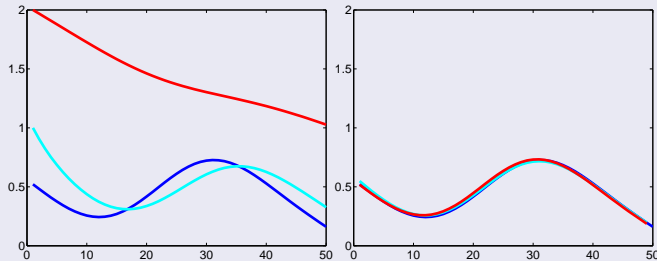


Figure: Left: joint samples from the transcription covariance, blue: $f(t)$, cyan: $x_1(t)$ and red: $x_2(t)$. Right: numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

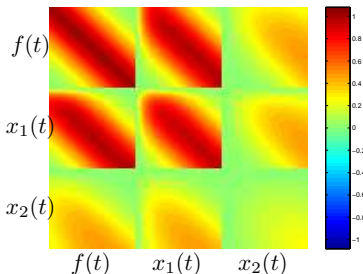
Covariance for Transcription Model

RBF Kernel function for $f(t)$

$$x_i(t) = \frac{B_i}{D_i} + S_i \exp(-D_i t) \int_0^t f(u) \exp(D_i u) du.$$

- Joint distribution for $x_1(t)$, $x_2(t)$ and $f(t)$.
- Here:

D_1	S_1	D_2	S_2
5	5	0.5	0.5



Noise Corruption

Estimate Underlying Noise

- Allow the mRNA abundance of each gene at each time point to be corrupted by noise, for observations at t_i for $i = 1, \dots, T$,

$$y_j(t_i) = x_j(t_i) + \epsilon_j(t_i) \quad (5)$$

with $\epsilon_j(t_i) \sim \mathcal{N}(0, \sigma_{ji}^2)$.

- Estimate noise level using probe-level processing techniques of Affymetrix microarrays (e.g. mmgMOS, [Liu et al., 2005]).
- The covariance of the noisy process is then $K_{yy} = \Sigma + K_{xx}$, with $\Sigma = \text{diag}(\sigma_{11}^2, \dots, \sigma_{1T}^2, \dots, \sigma_{N1}^2, \dots, \sigma_{NT}^2)$.

Artificial Data

Toy Problem

- Results from an artificial data set.
- We used a 'known TFC' and derived six 'mRNA profiles'.
 - Known TFC composed of three Gaussian basis functions.
 - mRNA profiles derived analytically.
- Fourteen subsamples were taken and corrupted by noise.
- This 'data' was then used to infer a distribution over plausible TFCs.

Artificial Data Results

demToyProblem1

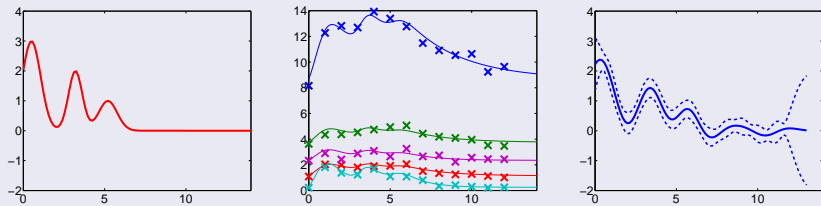


Figure: *Left:* The TFC, $f(t)$, which drives the system. *Middle:* Five gene mRNA concentration profiles each obtained by using different parameter sets $\{B_i, S_i, D_i\}_{i=1}^5$ (lines) along with noise corrupted 'data'. *Right:* The inferred TFC (with error bars).

Results

Linear System

- Recently published biological data set studied using linear response model by Barenco et al. [2006].
- Study focused on the tumour suppressor protein p53.
- mRNA abundance measured for five targets: *DDB2*, *p21*, *SESN1/hPA26*, *BIK* and *TNFRSF10b*.
- Quadratic interpolation for the mRNA production rates to obtain gradients.
- They used MCMC sampling to obtain estimates of the model parameters B_j , S_j , D_j and $f(t)$.

Linear response analysis

Experimental Setup

- We analysed data using the linear response model
- Raw data was processed using the mmgMOS model of Liu et al. [2005] which provides variance as well as expression level.
- We present posterior distribution over TFCs.
- Results of inference on the values of the hyperparameters B_j , S_j and D_j .
 - Samples from the posterior distribution were obtained using Hybrid Monte Carlo (see e.g. Neal, 1996).

Linear Response Results

demBarenco1

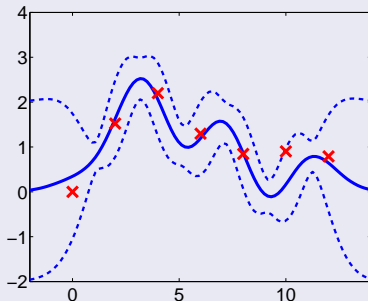


Figure: Predicted protein concentration for p53. Solid line is mean, dashed lines 95% credibility intervals. The prediction of [Barenco et al., 2006] was pointwise and is shown as crosses.

Results — Transcription Rates

Estimation of Equation Parameters demBarenco1

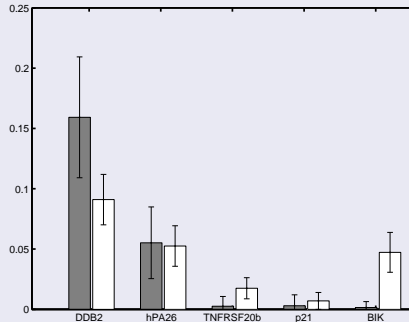


Figure: Basal transcription rates. Our results (black) compared with Barenco et al. [2006] (white).

Results — Transcription Rates

Estimation of Equation Parameters demBarenco1

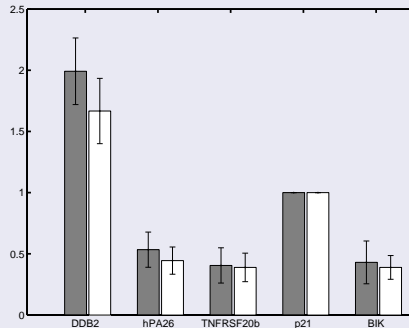


Figure: Sensitivities. Our results (black) compared with Barenco et al. [2006] (white).

Results — Transcription Rates

Estimation of Equation Parameters demBarenco1

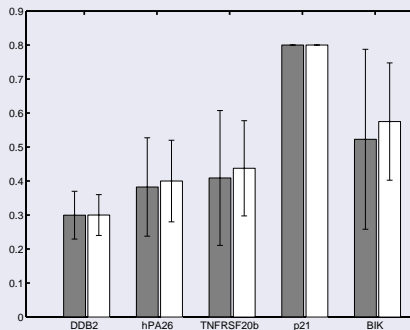


Figure: Decays. Our results (black) compared with Barenco et al. [2006] (white).

Linear Response Discussion

GP Results

- Note oscillatory behaviour, possible artifact of RBF covariance Rasmussen and Williams [see page 123 in 2006].
- Results are in good accordance with the results obtained by Barenco et al..
- Differences in estimates of the basal transcription rates probably due to:
 - different methods used for probe-level processing of the microarray data.
 - Our failure to constrain $f(0) = 0$.
- Our results take about 13 minutes to produce Barenco et al. required 10 million iterations of Monte Carlo.

Non-linear Response Model

More Realistic Response

- All the quantities in equation (1) are positive, but direct samples from a GP will not be.
- Linear models don't account for saturation.
- *Solution*: model response using a positive nonlinear function.

Formalism

Non-linear Response

- Introduce a non-linearity $g(\cdot)$ parameterised by θ_j

$$\frac{dx_j}{dt} = B_j + g(f(t), \theta_j) - D_j x_j$$
$$x_j(t) = \frac{B_j}{D_j} + \exp(-D_j t) \int_0^t du g(f(u), \theta_j) \exp(D_j u) .$$

- The induced distribution of $x_j(t)$ is no longer a GP.
- Derive the functional gradient and learn a MAP solution for $f(t)$.
- Also compute Hessian so we can approximate the marginal likelihood.

Implementation

Riemann quadrature

- Implementation requires a discretised time.
- Compute the gradient and Hessian on a grid.
- Integrate them by approximate Riemann quadrature.
- We choose a uniform grid $\{t_p\}_{p=1}^M$ so that $\Delta = t_p - t_{p-1}$ is constant.
- The vector $\mathbf{f} = \{f_p\}_{p=1}^M$ is the function f at the grid points.

$$I(t) = \int_0^t f(u) \exp(D_j u) du$$

$$I(t) \approx \sum_{p=1}^M f(t_p) \exp(D_j t_p) \Delta$$

Log Likelihood

Functional Gradient

- Given noise-corrupted data $y_j(t_i)$ the log-likelihood is

$$\log p(Y|f, \theta_j) = -\frac{1}{2} \sum_{i=1}^T \sum_{j=1}^N \left[\frac{(x_j(t_i) - y_j(t_i))^2}{\sigma_{ji}^2} - \log(\sigma_{ji}^2) \right] - \frac{NT}{2} \log(2\pi)$$

- The functional derivative of the log-likelihood wrt f is

$$\frac{\delta \log p(Y|f)}{\delta f(t)} = -\sum_{i=1}^T \Theta(t_i - t) \sum_{j=1}^N \frac{(x_j(t_i) - y_j(t_i))}{\sigma_{ji}^2} g'(f(t)) e^{-D_j(t_i - t)}$$

$\Theta(x)$ — Heaviside step function.

Log Likelihood

Functional Hessian

- Given noise-corrupted data $y_j(t_i)$ the log-likelihood is

$$\log p(Y|f, \theta_j) = -\frac{1}{2} \sum_{i=1}^T \sum_{j=1}^N \left[\frac{(x_j(t_i) - y_j(t_i))^2}{\sigma_{ji}^2} - \log(\sigma_{ji}^2) \right] - \frac{NT}{2} \log(2\pi)$$

- The negative Hessian of the log-likelihood wrt f is

$$\begin{aligned} w(t, t') &= \sum_{i=1}^T \Theta(t_i - t) \delta(t - t') \sum_{j=1}^N \frac{(x_j(t_i) - y_j(t_i))}{\sigma_{ji}^2} g''(f(t)) e^{-D_j(t_i - t)} \\ &+ \sum_{i=1}^T \Theta(t_i - t) \Theta(t_i - t') \sum_{j=1}^N \sigma_{ji}^{-2} g'(f(t)) g'(f(t')) e^{-D_j(2t_i - t - t')} \end{aligned}$$

$$g'(f) = \partial g / \partial f \text{ and } g''(f) = \partial^2 g / \partial f^2.$$

Implementation II

Combine with Prior

- Combine these with prior to compute gradient and Hessian of log posterior $\Psi(\mathbf{f}) = \log p(Y|\mathbf{f}) + \log p(\mathbf{f})$ [see Rasmussen and Williams, 2006, chapter 3]

$$\begin{aligned}\frac{\partial \Psi(\mathbf{f})}{\partial \mathbf{f}} &= \frac{\partial \log p(Y|\mathbf{f})}{\partial \mathbf{f}} - K^{-1}\mathbf{f} \\ \frac{\partial^2 \Psi(\mathbf{f})}{\partial \mathbf{f}^2} &= -(W + K^{-1})\end{aligned}\tag{6}$$

K prior covariance evaluated at the grid points.

- Use to find a MAP solution via, $\hat{\mathbf{f}}$, using Newton's algorithm.
- The Laplace approximation is then

$$\log p(Y) \simeq \log p(Y|\hat{\mathbf{f}}) - \frac{1}{2}\hat{\mathbf{f}}^T K^{-1}\hat{\mathbf{f}} - \frac{1}{2} \log |I + KW|.\tag{7}$$

Example: linear response

Using non-RBF kernels

- Start by taking $g(\cdot)$ to be linear.
- Provides 'sanity check' and smooth (*i.e.* non-stochastic) covariance function.
- Avoids double numerical integral that would normally be required.

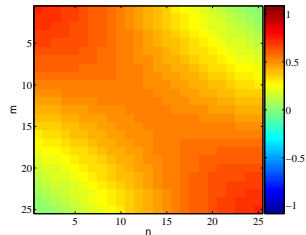
Oscillatory Behaviour

Fix with MLP Covariance

MLP Kernel Function

$$k(t, t') = \alpha \arcsin \left(\frac{w t t' + b}{\sqrt{(w t^2 + b + 1)(w t'^2 + b + 1)}} \right)$$

- A non-stationary covariance matrix [Williams, 1997].
- Derived from a multi-layer perceptron (MLP).



Covariance Samples

demCovFuncSample (oxford toolbox)

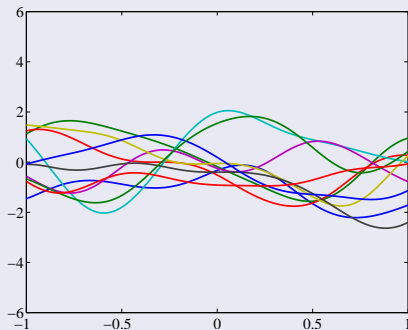


Figure: RBF kernel with $l = 0.25$, $\alpha = 1$

Covariance Samples

demCovFuncSample (oxford toolbox)

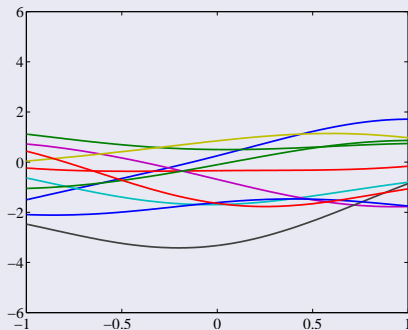


Figure: RBF kernel with $l = 1$, $\alpha = 1$

Covariance Samples

demCovFuncSample (oxford toolbox)

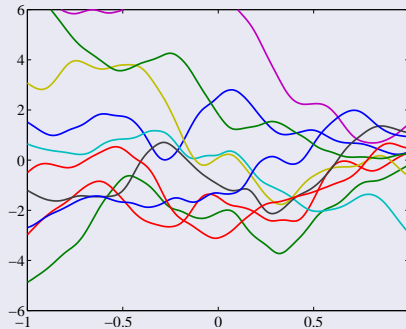


Figure: MLP kernel with $\alpha = 8$, $w = 100$ and $b = 100$

Covariance Samples

demCovFuncSample (oxford toolbox)

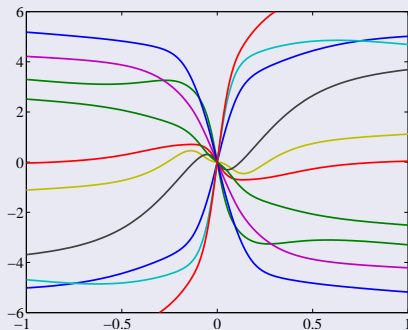


Figure: MLP kernel with $\alpha = 8$, $b = 0$ and $w = 100$

Response Results

demBarencoMap1, demBarencoMap2

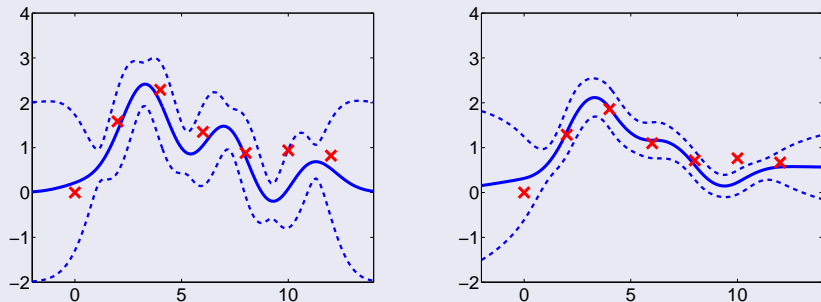


Figure: Predicted protein concentration for p53 using a MAP estimation with linear response model: *Left:* RBF prior on f (log likelihood -101.4); *Right:* MLP prior on f (log likelihood -105.6). Solid line is mean prediction, dashed lines are 95% credibility intervals. The prediction of Barenco *et al.* was pointwise and is shown as crosses.

Non-linear response analysis

Non-linear responses

- Exponential response model (constrains protein concentrations positive).
- $\log(1 + \exp(f))$ response model.
- $\frac{3}{1 + \exp(-f)}$
- Inferred MAP solutions for the latent function f are plotted below.

$\exp(\cdot)$ Response Results

demBarencoMap3, demBarencoMap4

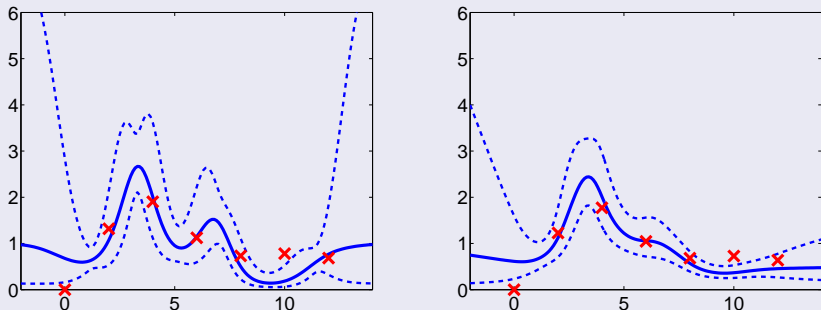


Figure: Predicted protein concentration for p53 using an exponential response: *Left:* shows results of using a squared exponential prior covariance on f (log likelihood -100.6); *Right:* shows results of using an MLP prior covariance on f (log likelihood -106.4). Solid line is mean prediction, dashed lines show 95% credibility intervals. The results shown are for $\exp(f)$, hence the asymmetry of

$\log(1 + \exp(f))$ Response Results

demBarencoMap5, demBarencoMap6

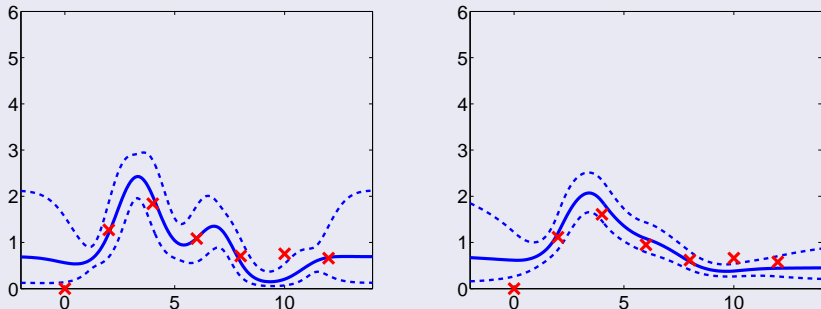


Figure: Predicted protein concentration for p53 using a $\log(1 + \exp(x))$ response: *Left:* shows results of using a squared exponential prior covariance on f (log likelihood -100.9); *Right:* shows results of using an MLP prior covariance on f (log likelihood -110.0). Solid line is mean prediction, dashed lines show 95% credibility intervals. The results shown are for $\exp(f)$, hence the

$\frac{3}{1+\exp(-f)}$ Response Results

demBarencoMap7, demBarencoMap8

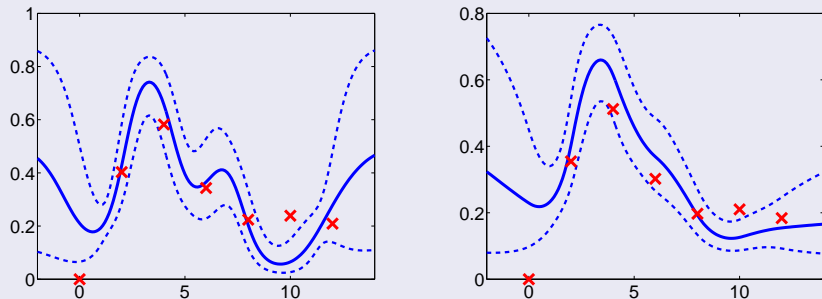


Figure: Predicted protein concentration for p53 using a sigmoid response: *Left:* shows results of using a squared exponential prior covariance on f (log likelihood -104.1); *Right:* shows results of using an MLP prior covariance on f (log likelihood -111.2). Solid line is mean prediction, dashed lines show 95% credibility intervals. The results shown are for $\exp(f)$, hence the asymmetry of

Discussion

- We have described how GPs can be used in modelling dynamics of a simple regulatory network motif.
- Our approach has advantages over standard parametric approaches:
 - there is no need to restrict the inference to the observed time points, the temporal continuity of the inferred functions is accounted for naturally.
 - GPs allow us to handle uncertainty in a natural way.
 - MCMC parameter estimation in a discretised model can be computationally expensive. Parameter estimation can be achieved easily in our framework by type II maximum likelihood or by using efficient hybrid Monte Carlo sampling techniques
- All code on-line
<http://www.dcs.shef.ac.uk/~neil/gpsim/>.

Future Directions

- This is still a very simple modelling situation.
 - We are ignoring transcriptional delays.
 - Here we have single transcription factor: our ultimate goal is to describe regulatory pathways with more genes.
 - All these issues can be dealt with in the general framework we have described.
 - Need to overcome the greater computational difficulties.

Other Relevant Work

- Simpler genome wide models [Sabatti and James, 2006, Sanguinetti et al., 2006b,a].
- Better MCMC models [Rogers et al., 2006].
- Sorry for anyone missed!

Open Question

Functional Approximations

- Without sampling, how can we produce a better posterior approximation?

Acknowledgements

Data and Support

We thank Martino Barenco for useful discussions and for providing the data. We gratefully acknowledge support from BBSRC Grant No BBS/B/0076X “Improved processing of microarray data with probabilistic models”.

References I

- M. Barenco, D. Tomescu, D. Brewer, R. Callard, J. Stark, and M. Hubank. Ranked prediction of p53 targets using hidden variable dynamic modeling. *Genome Biology*, 7(3):R25, 2006.
- T. Graepel. Solving noisy linear operator equations by Gaussian processes: Application to ordinary and partial differential equations. In T. Fawcett and N. Mishra, editors, *Proceedings of the International Conference in Machine Learning*, volume 20, pages 234–241. AAAI Press, 2003. ISBN 1-57735-189-4.
- X. Liu, M. Milo, N. D. Lawrence, and M. Rattray. A tractable probabilistic model for Affymetrix probe-level analysis across multiple chips. *Bioinformatics*, 21(18):3637–3644, 2005.
- R. Murray-Smith and B. A. Pearlmutter. Transformations of Gaussian process priors.
- R. M. Neal. *Bayesian Learning for Neural Networks*. Springer, 1996. Lecture Notes in Statistics 118.
- A. O'Hagan. Curve fitting and optimal design for prediction. *Journal of the Royal Statistical Society, B*, 40:1–42, 1978.
- A. O'Hagan. Some Bayesian numerical analysis. In J. M. Bernardo, J. O. Berger, A. P. Dawid, and A. F. M. Smith, editors, *Bayesian Statistics 4*, pages 345–363, Valencia, 1992. Oxford University Press.
- C. E. Rasmussen and C. K. I. Williams. *Gaussian Processes for Machine Learning*. MIT Press, Cambridge, MA, 2006. ISBN 026218253X.
- S. Rogers, R. Khanin, and M. Girolami. Model based identification of transcription factor activity from microarray data. In *Probabilistic Modeling and Machine Learning in Structural and Systems Biology*, Tuusula, Finland, 17-18th June 2006.
- C. Sabatti and G. M. James. Bayesian sparse hidden components analysis for transcription regulation networks. *Bioinformatics*, 22(6):739–746, 2006.
- G. Sanguinetti, N. D. Lawrence, and M. Rattray. Probabilistic inference of transcription factor concentrations and gene-specific regulatory activities. *Bioinformatics*, pages 2275–2281, 2006a. Advance Access.
- G. Sanguinetti, M. Rattray, and N. D. Lawrence. A probabilistic dynamical model for quantitative inference of the regulatory mechanism of transcription. *Bioinformatics*, 22(14):1753–1759, 2006b.
- C. K. I. Williams. Computing with infinite networks. In M. C. Mozer, M. I. Jordan, and T. Petsche, editors, *Advances in Neural Information Processing Systems*, volume 9, Cambridge, MA, 1997. MIT Press.