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# **Aurora-2 User's Manual**

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April, 2000

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# Preface

## INTRODUCTION

The Aurora-2 is a Near-field Scanning Optical Microscope (NSOM). NSOM is an optical microscopy technique that offers higher resolution limits than confocal microscopy. Because NSOM combines optical and scanning probe microscopy, sample surface chemistry can be imaged and analyzed simultaneously with surface topography measurements. The ability to gather topographic data while producing an optical image is useful in material characterization and analysis.

It is important to read this manual and be familiar with the Aurora-2 instrument to facilitate more productive and efficient use of NSOM as a research tool.

## ABOUT THIS MANUAL

Chapter 1 provides an overview of the theory of NSOM and how it is applied in the Aurora-2 instrument. Chapter 2 describes the Aurora-2 instrument components and set-up. Chapters 3 and 4 provide step-by-step instructions for taking a topographic scan and an NSOM scan, respectively, of the standard sample.

This manual covers those aspects of the SPMLab software that are specific to the Aurora-2 configuration. For a thorough understanding of how to use the software, the user should refer to the *SPMLab Software Reference Manual*.

## OPERATING SAFETY

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All Warning and Caution statements in this manual should be strictly observed. Failure to do so may result in serious injury, particularly blindness due to exposure to laser light, and damage to your Aurora-2 instrument.



Warning statements—indicated by this symbol—alert you to possible serious injury, especially blindness due to laser light exposure. Procedures in this manual must be followed exactly. Do not proceed beyond a warning until the conditions of the warning are understood and met.



Caution statements—indicated by this symbol—call attention to possible damage to the system or to the impairment of safety unless procedures described in this manual are followed exactly.

---

**WARNING**  ***NEVER LOOK DIRECTLY INTO THE LASER BEAM.***

---



These labels are placed on the Aurora-2 to warn you of the danger of laser light. Exposure to laser light is possible at the laser, at the end of the laser fiber, at the probe tip, or at any point on the fiber-optic cable if it should break. Follow all set-up and operation procedures in this manual carefully.

# THERMOMICROSCOPES PRODUCT WARRANTY

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## COVERAGE

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Instruments, parts, and accessories not manufactured by ThermoMicroscopes may be warranted by ThermoMicroscopes for the specific items and periods expressed in writing on published price lists or quotes. However, all such warranties extended by ThermoMicroscopes are limited in accordance with all the terms, conditions, and other provisions stated in this warranty. ThermoMicroscopes makes no warranty whatsoever concerning products or accessories not of its manufacture except as noted above.

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4. Provide adequate and safe working space around the products for servicing by ThermoMicroscopes personnel.

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## WARRANTY LIMITATIONS

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been modified or altered without written authorization from ThermoMicroscopes; *f*) Products which have had the serial number altered or removed; *g*) Improper or inadequate care, maintenance, adjustment, or calibration by the user.

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# Chapter 1

## Theory of Operation

### NSOM THEORY

Historically, the limiting factor in optical microscopy has been the diffraction limit of light. Attempts to image features approximately the size of a wavelength of light met with frustration because of diffraction. As far back as the 1930's, a theoretical solution to this problem was suggested.<sup>1</sup> However, it was not until the early 1980's that the electronic control and feedback capability existed to realize this solution.<sup>2</sup> If the aperture exposing the sample is kept very small—on the order of 50 nm—and the aperture is kept close to the sample surface—generally less than 20 nm—the diffraction problems can be avoided. It was not until the development of scanning probe microscopes, however, that technology existed to maintain such close tip-sample spacing while a tip was being scanned over a sample.

### NSOM TIPS

The light source in an NSOM system is launched into an optical fiber. The end of the fiber is “pulled down” to a diameter of 50 nm. The fiber is then coated with aluminum, approximately 100 nm thick. The fiber becomes a “light funnel” directing light onto the sample. Photodetectors are placed behind the sample (transmission mode) or beside the tip (reflection mode) to collect light emitted from the sample. A laser is used as a light source and is coupled into the back side of the NSOM fiber-optic probe.

1. E.H. Synge: A Suggested Model for Extending Microscopic Resolution into the Ultramicroscopic Region. *Phil. Mag.* 6, 356-362 (1928).

2. D.W. Pohl, W. Denk, and M. Lanz: Optical Stethoscopy: Image Recording with Resolution 1/20. *Appl. Phys. Lett.* 44, No. 7, 651-653 (1984).

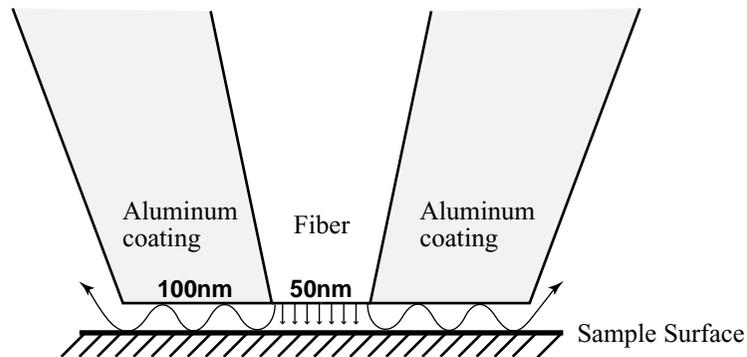


Figure 1-1 Fiber-Optic Probe Tip

## DISTANCE CONTROL MECHANISMS

Tip geometry alone is not enough to produce images. The tip must also be very close to the sample, typically  $<10$  nm. These small separations are only possible with electronic position detection and control. Distance control is currently achieved using a shear-force mechanism. Two such mechanisms exist: **light-lever** and **tuning fork**. For these techniques, the tip is attached to a vibrating element which is driven at its resonant frequency. This vibration is parallel to the surface. As the tip approaches the surface, the vibration amplitude and phase change. This change in amplitude and phase generates an electrical signal that is input into the feedback loop.

The Aurora-2 uses the tuning fork mechanism for distance control, as it has the advantage of producing an electrical signal directly, rather than relying on another device to generate a signal. This direct connection provides better feedback control, uses much smaller vibration amplitudes, and does not introduce unwanted light into the sample area.

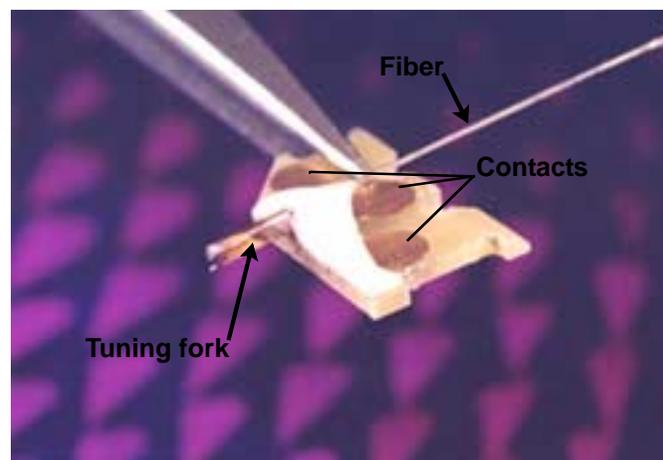


Figure 1-2 Aurora-2 Fiber-Optic Probe

The fiber-optic probe is attached to one prong of a piezoelectric tuning fork. The tip extends slightly beyond the end of the tuning fork. The tuning fork is vibrated using a dithering piezoelectric device, which produces a vibrational amplitude at the tip of approximately one nanometer. As the piezoelectric material of the fork vibrates, it produces a small current. The vibration amplitude of the fiber can then be measured by measuring the piezoelectric signal from the tuning fork. The current is amplified and input into a lock-in amplifier. The phase change of the tuning fork signal relative to the driving signal is measured and used in the feedback loop.

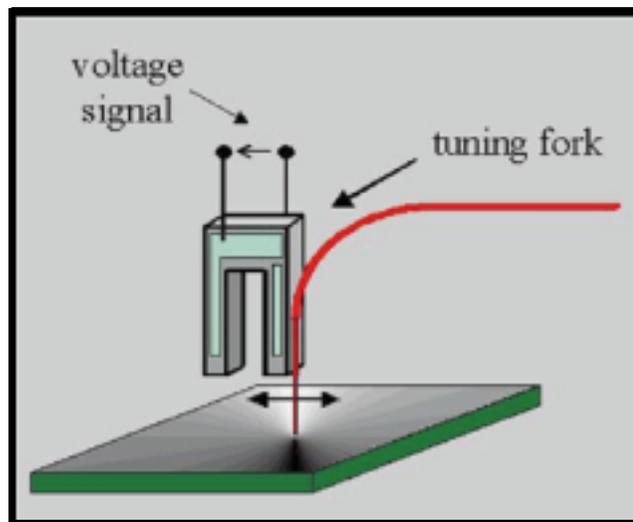


Figure 1-3 Tuning Fork Mechanism

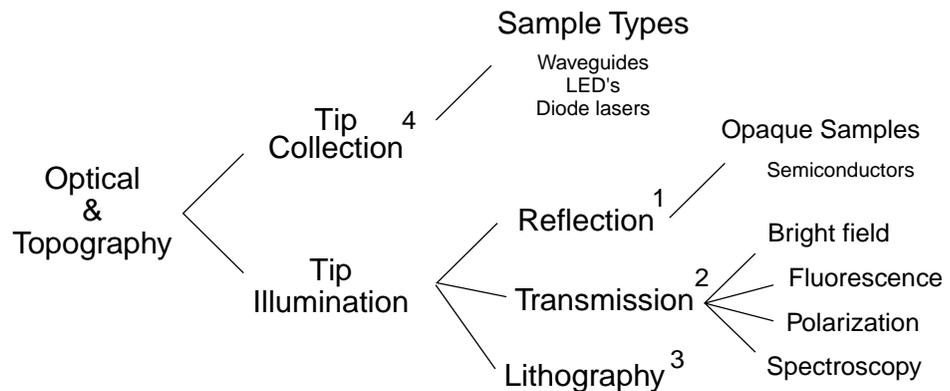
## MODES OF OPERATION

The NSOM instrument is a unique combination of a scanning probe microscope and an ultra-high resolution optical microscope. Generally both modes—topography and optical—are used together. However, there may be some situations where topography is used alone.

The NSOM, used as an optical microscope, can be operated either in **tip collection** or **tip illumination** mode. In tip collection mode, the sample is the light source, and the tip acts as a way to collect this light. This method is best for samples such as waveguides and laser diodes.

Tip illumination mode is perhaps the most common NSOM mode. Tip illumination uses the tip as a “light funnel” to illuminate the sample in a precise, controlled manner. This mode can be further subdivided into **reflection** collection, **transmission** collection, and **lithography** modes. Reflection collection gathers light that has been reflected from the sample. It is used for opaque samples, such as semiconductor materials. This method is not as efficient for gathering light, since the physical position of the tip collector does not allow it to collect much of the reflected light. Reflection mode might also suffer from image artifacts created by tip shadowing on the sample surface. Transmission mode is more commonly used and is more efficient. The collector is placed behind the sample and collects a majority of the light as it passes through the sample. The drawback to this mode is that it requires the use of thin, transparent samples.

There are a number of operational techniques in transmission mode. The major techniques are **bright field**, **fluorescence**, **polarization**, and **spectroscopy**. These techniques use different properties of light. Bright field mode is similar to standard optical microscopy in that the sample is exposed to light from the NSOM probe, and the resulting image is recorded by detecting all wavelengths, including the source light, on the photomultiplier tube (PMT). In fluorescence modes, the tip is used to excite the sample, and any resulting fluorescence is captured and imaged. Polarization mode typically polarizes the incoming light and looks at how the sample changes that polarization. In spectroscopy techniques, the signal is the change (either time-scale or wavelength) the sample causes in the exposing light. Figure 1-4 illustrates the relationships between these operational modes.



**Figure 1-4 NSOM Operational Modes**

Examples of NSOM imaging with the Aurora, using the modes referenced in Figure 1-4, are included in the following literature.

1. P.J. Moyer, T. Cloninger, J. Gole, and L. Bottomley. Experimental evidence for molecule-like absorption and emission of porous silicon using near-field and far-field optical spectroscopy. *Phys. Rev. B*, 60, No. 7, 4889-4896 (1999).
2. P.F. Barbara, D.M. Adams, and D.B. O'Connor. Characterization of organic thin film materials with near-field scanning optical microscopy (NSOM). *Annu. Rev. Mater. Sci.*, 29, 433-469 (1999).
3. A. Naber, H. Kock, and H. Fuchs. High-Resolution Lithography with Near-Field Optical Microscopy. *Scanning* Vol. 18 (8), 567-571 (1996).
4. Ch. Lienau, A. Richter, A. Klehr, and T. Elsaesser. Near-Field Scanning Optical Microscopy of Polarization Bistable Laser Diodes. *Appl. Phys. Lett.* 69, No. 17, 2471-2473 (1996).

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## Chapter 2

# Aurora-2 Overview & Set-Up

### INSTRUMENT OVERVIEW

The Aurora-2 instrument is a platform for obtaining topographic and optical images. It offers a wide variety of configuration options, depending on the desired NSOM imaging mode. The sample is mounted on a scanning stage which is controlled by a three-piezo scanner arrangement. The fiber-optic probe is mounted on the removable Aurora-2 microscope head and positioned above the sample. Topographic and optical images can be taken simultaneously.

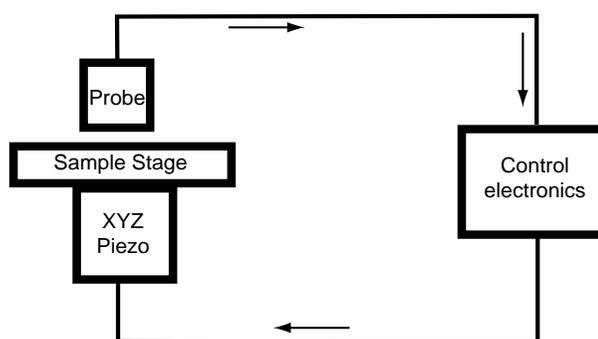


Figure 2-1 Topography Feedback Loop

The optical components of the Aurora-2 system are used for taking NSOM data as well as for focusing the optics and monitoring the probe-sample approach. The rotating mirror (see Figure 2-2) selects either the reflection or transmission objective. The two “flipper” mirrors can be manually flipped down to allow the use of the PMT or optional hardware, such as a photon counter or spectrometer.

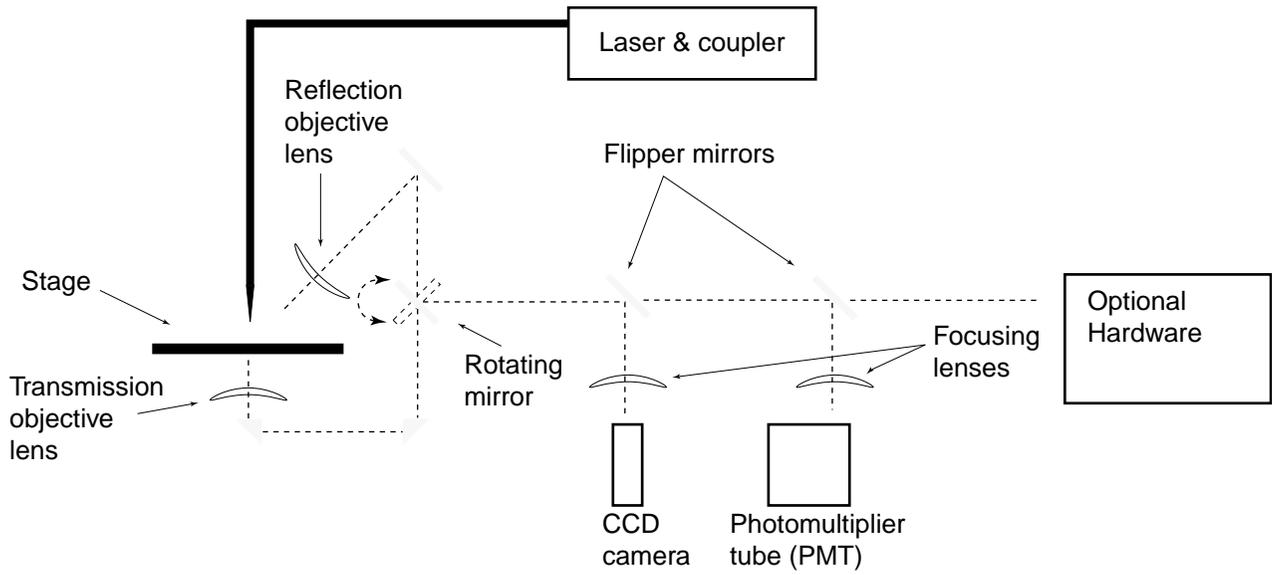


Figure 2-2 Aurora-2 Light Path

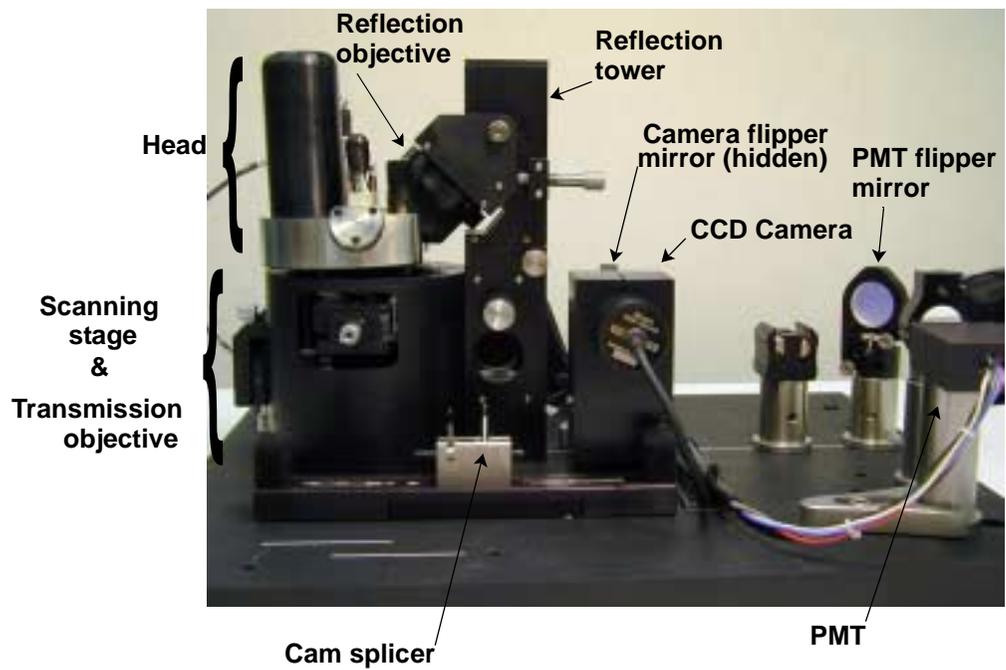


Figure 2-3 Instrument Components

## AURORA-2 PACKAGE

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### STANDARD COMPONENTS

All the components of the basic Aurora-2 configuration are listed below. It is a good idea to go through this list to be sure that all the items have been received. Shipping errors can be corrected by contacting Customer Service.

- Instrument stage (base plate with mounted hardware)
- Aurora-2 sensor head
- Electronic Control Unit-*Plus* (ECU-*Plus*) with I/O 10 and I/O MOD+ boards
- Aurora Control Unit
- Computer
- Video monitor
- NSOM fiber-optic tips
- Probe installation tool
- Fiber cleaver
- Fiber stripper tool
- Tool kit
- Cables
- NSOM standard sample
- User's Manual
- Instrument enclosure
- SPMLab software
- *SPMLab Software Reference Manual*

### OPTIONAL EQUIPMENT

- I/O-U input/output board
- I/O-P photon counter board
- Laser
- Laser coupler
- Daughter board for additional analog-to-digital conversion channels
- Explorer SPM head
- Vibration isolation table

The Explorer SPM head is a popular option, as it uses the same control hardware as the Aurora-2. The User-Access board (I/O-U) provides access to most of the input and monitor signals on the ECU-*Plus*. The photon counter interface board (I/O-P) allows the user to collect data through a photon counter, which is useful for very low light levels. Contact the ThermoMicroscopes representative in your area for more information on these options.



Figure 2-4 Probe and Fiber Tools

## INSTALLATION

### INSTRUMENT LOCATION CONSIDERATIONS

The Aurora-2 should be mounted in an environment that is as vibration-free as possible. Sources of mechanical and acoustic vibration will decrease the Aurora's maximum resolution capability. The Aurora-2 should be placed on a its own vibration isolation table. Computer cooling fans and mouse clicks produce vibration that negatively impacts image quality, so place the computer on a separate table. Basement or ground floor rooms are better for the instrument, since multi-story buildings usually have significant vibration on the upper floors. Temperature and humidity should be controlled to maintain constant environmental conditions. Normal indoor conditions, i.e., "room temperature" and average humidity, are sufficient. Extremes of temperature and humidity will negatively affect the instrument and possibly cause damage.

### CABLE CONNECTIONS

**CAUTION** ⚠ *Make sure the power is OFF to all the modules and the computer while setting up. Connecting cables to powered-up electronics may damage the modules.*

The cable connection diagrams (Figure 2-5 and Figure 2-6) show the configuration with and without an external lock-in amplifier (a lock-in amplifier is also integrated into the I/O MOD+ board). A photocopy of the appropriate diagram is useful for checking off the cable connections as they are made.

## POWERING UP THE SYSTEM

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**WARNING**  *To prevent serious injury, make sure the cover is on the laser coupler before turning the laser on. Follow all safety warnings when powering up and using the laser.*

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**CAUTION**  *Make sure the PMT voltage is turned all the way down (to the counter-clockwise limit) before powering up the components.*

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Once all the connections are made, the components can be powered up. First turn on the ECU-Plus, then the computer and the video monitor. (The ECU-Plus should always be turned on before the computer so that the ECU interface is recognized and initialized.) The laser is powered up separately. Power is automatically applied to the Aurora Control Unit when the ECU-Plus is powered up.

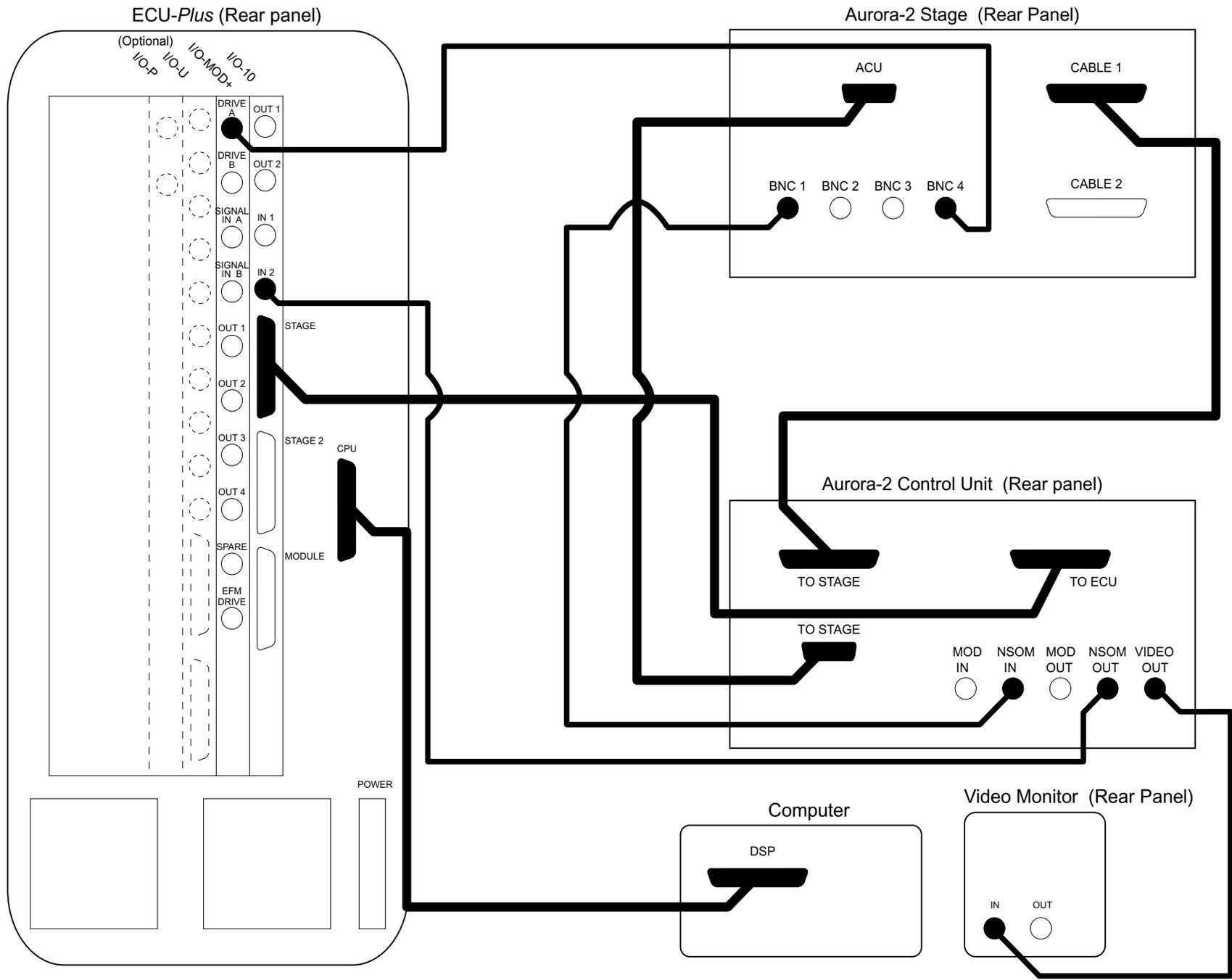


Figure 2-5 Aurora-2 Wiring Diagram

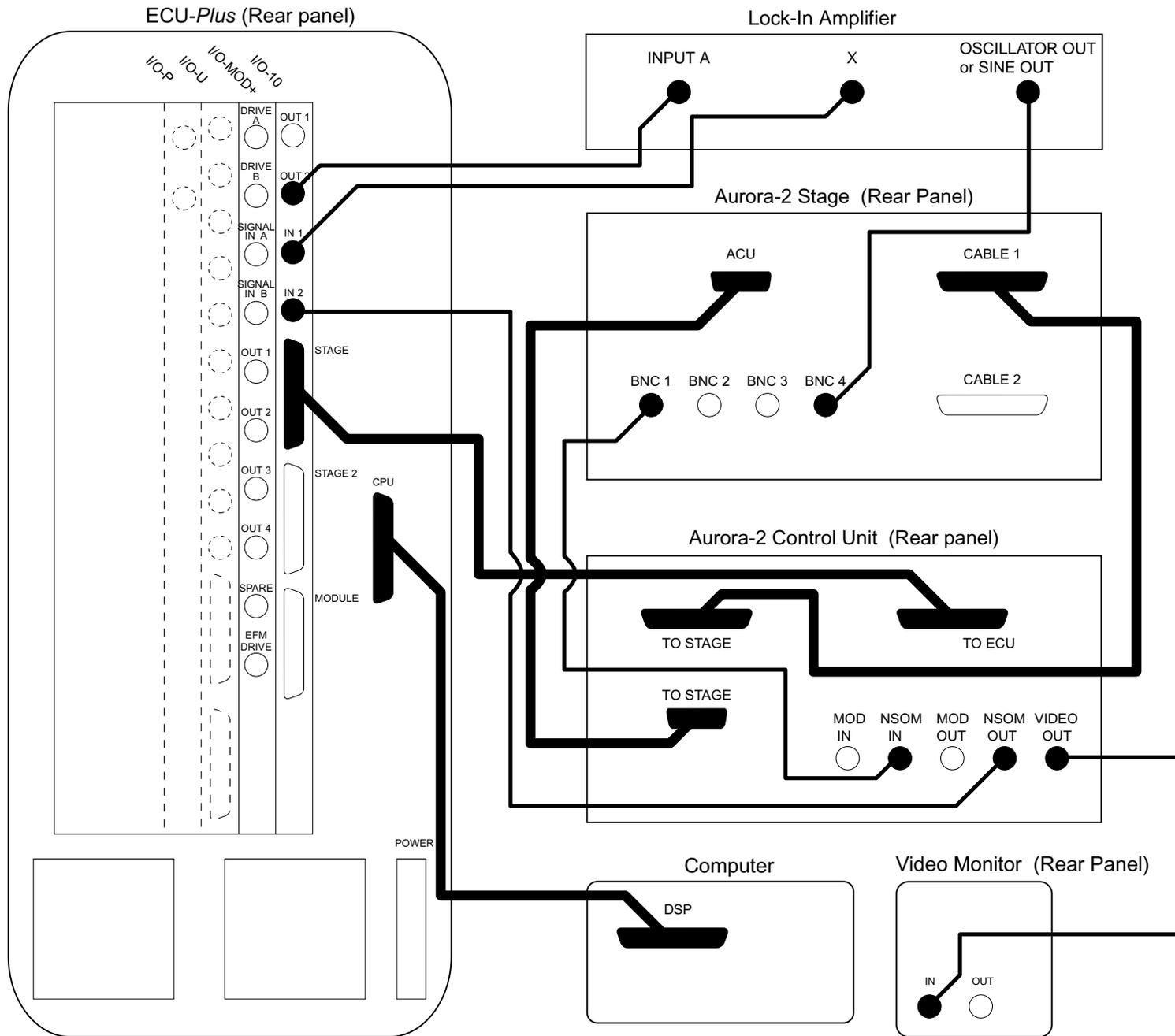
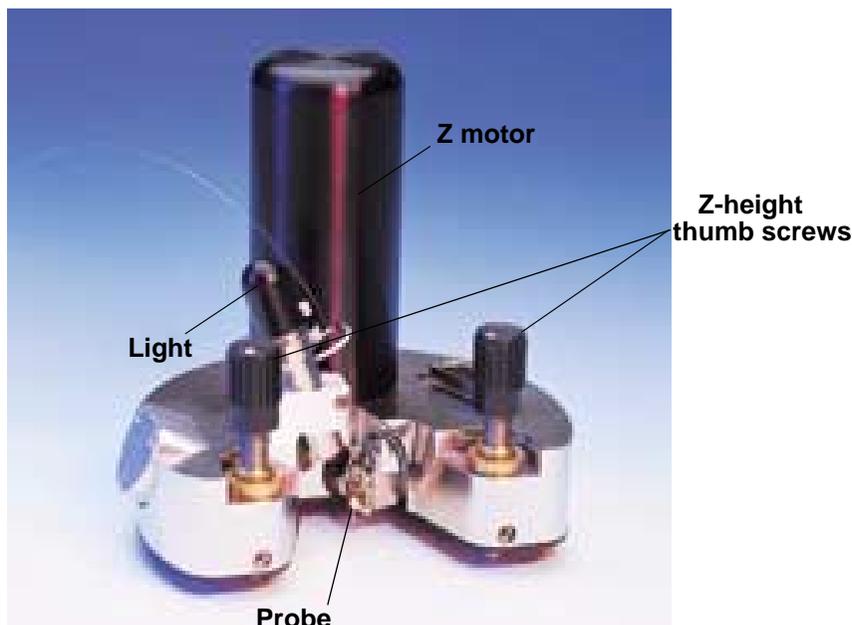


Figure 2-6 Aurora-2 Wiring Diagram for External Lock-in Amplifier

## THE AURORA-2 HEAD

**CAUTION** ⚠ *To avoid damage to the probe and sample, handle the head with care, paying particular attention to the probe and probe mount on the bottom. Whenever the head is removed from the microscope stage, use the Tip Up button  on the Data Acquisition tool bar to raise the tip a safe distance away from the sample. When placing the head on the stage, or when carefully setting it down on a table or other flat surface, turn the Z-height thumbscrews one full turn clockwise. Whenever the head is lowered using the Z motor or thumbscrews, watch the image on the video monitor to make sure the probe does not crash into the sample or stage.*



**Figure 2-7** Aurora-2 Head

The Aurora-2 head rests on a kinematic mount on the instrument base, as shown in Figure 2-8 (note that there is also a kinematic mount for the Explorer head). The Z-height between the probe and the sample stage is controlled by the three Z-height screws, as shown in Figure 2-9. On the bottom of each screw is a ball bearing that fits into the kinematic mount. The motorized screw (Z motor) is controlled with the Tip Up  and Tip Down  buttons on the Data Acquisition tool bar in the SPMLab software. The other two screws are adjusted manually with the thumbscrews on the head.

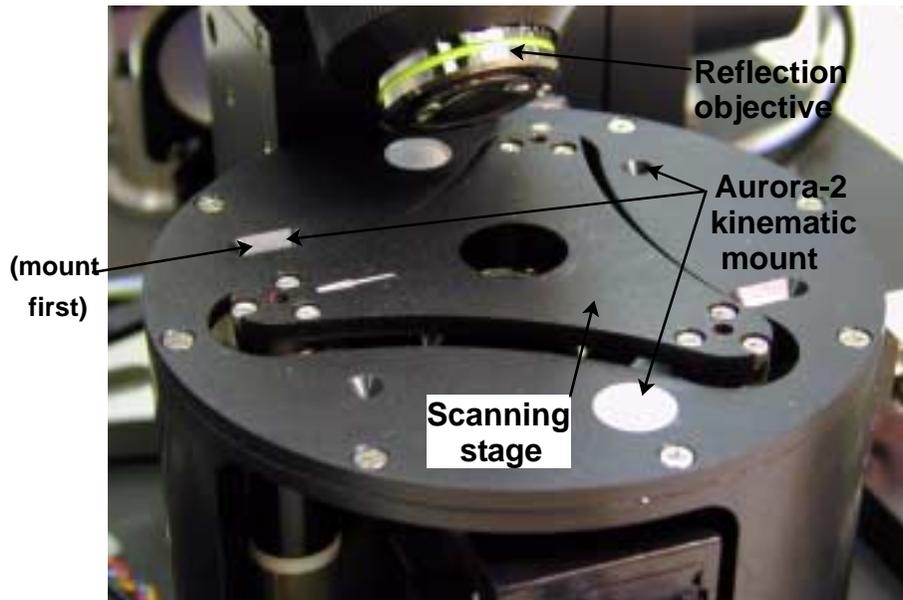


Figure 2-8 Kinematic Mount

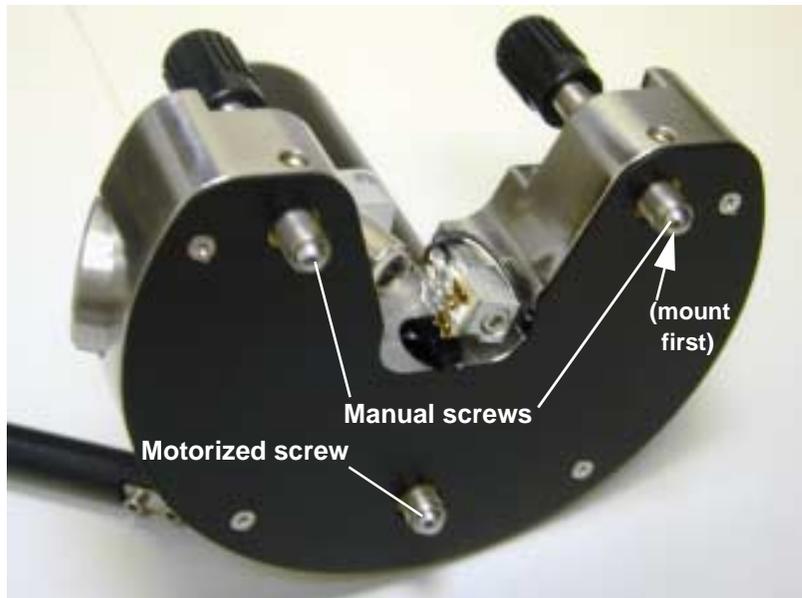


Figure 2-9 Z-Height Screws

## PLACING THE HEAD ON THE STAGE

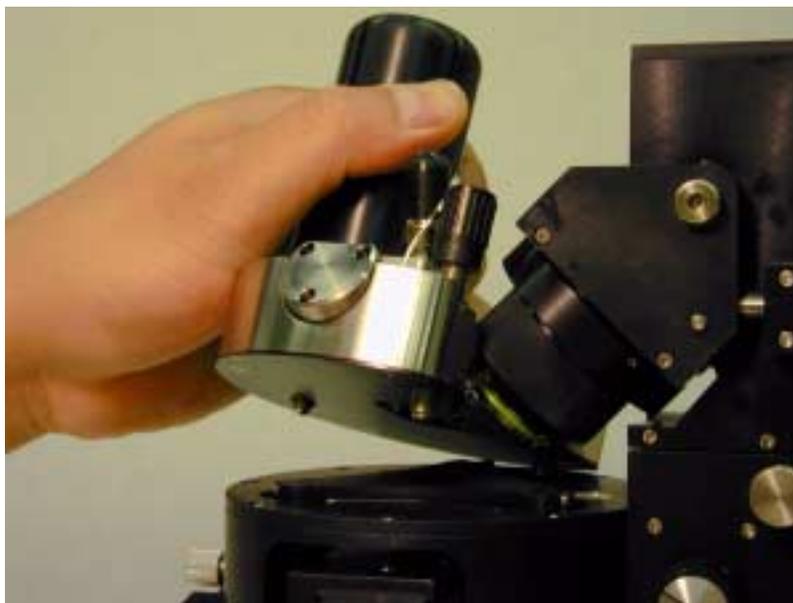
- Step 1** Turn the Z-height thumbscrews one full turn clockwise.
- Step 2** Connect the data cable to the female connector on the head.
- Step 3** Hold the head in your left hand, and position it over the stage, being careful not to hit the reflection objective.

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**CAUTION**  *To avoid damaging the piezo scanners, be careful not to touch the stage with the head.*

---

- Step 4** Check to see that there is sufficient clearance between the head and the scanning stage before lowering the head all the way onto the mount. If necessary, adjust the thumbscrews so the head does not end up resting on the stage instead of the kinematic mount.
- Step 5** Lower the head slowly onto the kinematic mount, tilting it slightly away from you so the manual Z-height screw opposite you lowers into the mount first, as shown in Figure 2-10.



**Figure 2-10** Placing the Head on the Stage

- Step 6** Lower the head onto the stage so the other two screws come to rest in the mount.
- Step 7** If a probe is mounted on the head, tape the fiber-optic cable to the Z motor enclosure on the head, as shown below in Figure 2-15 on page 14 under “Installing a Probe.”

## REMOVING THE HEAD

- Step 1** Click the Tip Up button  on the Data Acquisition tool bar to raise the tip a safe distance away from the sample.
- Step 2** Remove the head by first tilting it slightly away from you, pivoting it on the kinematic mounting, then lifting it safely off the stage.
- Step 3** Set the head down on a soft, flat surface, making sure there is no obstruction that would strike the probe.

## MOUNTING THE SAMPLE ON THE STAGE

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The sample included with the instrument was made by coating a surface with latex spheres approximately 450 nm in diameter, then depositing aluminum onto the surface. The spheres are then removed, leaving the coating on the interstitial areas, as shown in Figure 2-11.

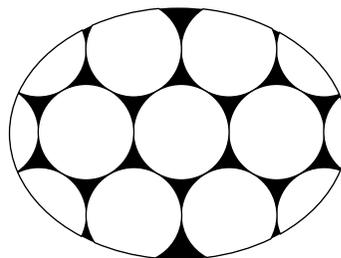


Figure 2-11 Standard Sample Example

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**CAUTION**  To avoid damage to the piezo scanners, do not apply more force than necessary to the scanning stage.

---

To mount the sample, you must remove the head. The sample slide can simply be scotch-taped to the stage. A piece of tape over each corner will hold the sample in place during the scan. This technique is the most common one for mounting samples on the Aurora-2 stage. Alternatively, small amounts of immersion oil can be used to hold the sample to the scanner plate.

## INSTALLING A PROBE

---

**CAUTION** ⚠ *Handle the probe with care, as it can be easily damaged.*

**Step 1** To grasp the probe with the probe installation tool, position the tool over the probe and fit the two “teeth” into the corresponding gaps on either side of the probe cartridge, as shown in Figure 2-12.



**Figure 2-12 Grasping the Probe with the Probe Tool**

- Step 2** Press down firmly into the foam lining of the probe box until the tool snaps into place.
- Step 3** Carefully pull the tape off the optical fiber so the probe can be removed from the box. Unroll the fiber, taking care not to touch the tuning fork or put any strain on the fiber.
- Step 4** Slide the probe into the holder in the head by sliding it under the clips, as shown in Figure 2-13.
- Step 5** Make sure the three clips make contact with the three metal contact pads on the probe, as shown in Figure 2-14. If necessary, use both thumbs to gently push the probe all the way into place, while cradling the head with both hands.



Figure 2-13 Sliding the Probe into the Holder

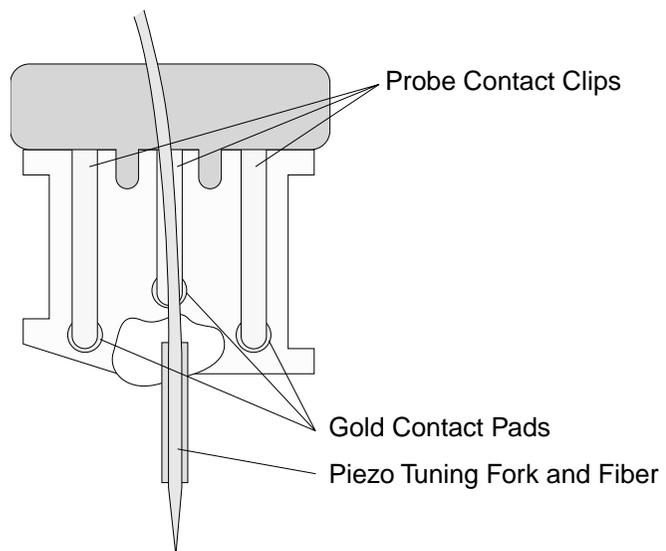


Figure 2-14 Probe Cartridge

- Step 6** Raise the thumbscrews located on the front of the head by turning them clockwise one full turn. This will keep the tip from hitting the stage when the head is placed on the stage.
- Step 7** Carefully place the head on the stage, and plug the data cable into the head, as described above on page 2-10.
- Step 8** Tape the fiber to the Z motor enclosure on the head, as shown in Figure 2-15. This takes strain off the fiber and helps prevent separation from the probe.



**Figure 2-15 Fiber Taped to the Aurora-2 Head**

- Step 9** Use the fiber stripper tool to strip away about 25mm of the polymer buffer off the end of the tip fiber. Note that once stripped, the fiber is very brittle.
- Step 10** Use the optical fiber cleaver from the tool kit to cleave the end of the tip fiber:
- Open the jaws of the cleaver, and seat the fiber into the notch in the lower jaw.
  - Insert the fiber until the stripped portion on the end aligns with the 16 mm mark.
  - Press the scoring bar down on the fiber. Take care not to break the fiber.
  - Gently flex the tongue of the cleaver down until the fiber cleaves. Take care not to touch the tip.
- Step 11** Carefully insert the fiber into one end of the cam splicer. Gently roll or twist the fiber as it is inserted to help it enter. Insert about 15mm into the splicer. Leave the coupler open, with the lock arms up.
- Step 12** Gently seat the fiber into the fiber holder on the end of the cam splicer assembly.

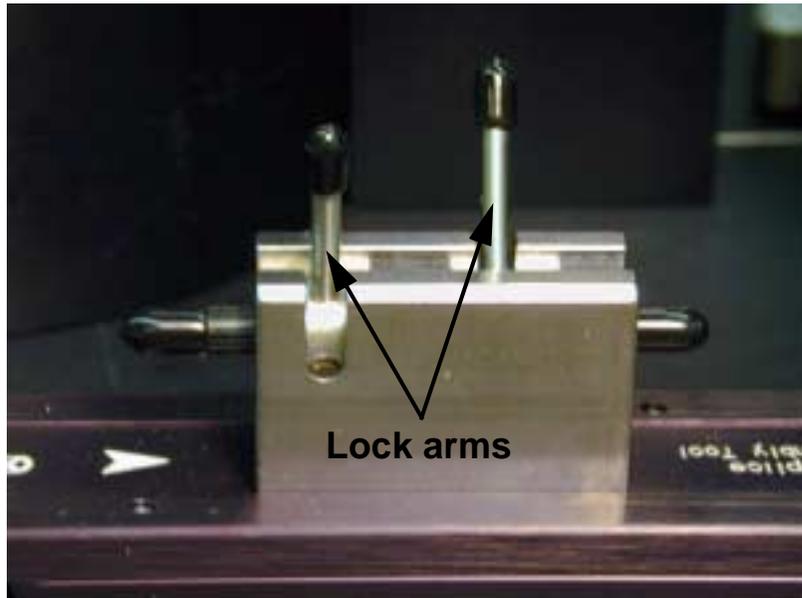


Figure 2-16 Cam Splicer

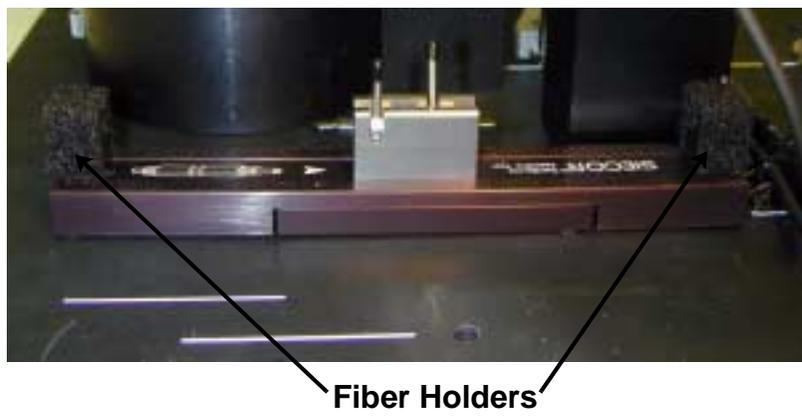
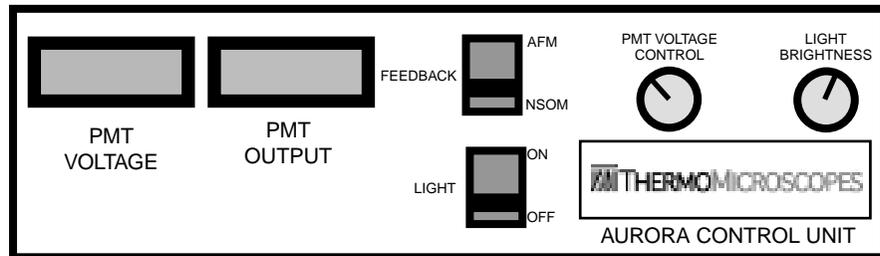


Figure 2-17 Cam Splicer Assembly

## AURORA CONTROL UNIT

**CAUTION**  *Keep the PMT Voltage on the Aurora Control Unit set to 0, unless the PMT is shielded from all sources of light (except the laser during NSOM imaging). Exposure to light can damage the PMT. The voltage should only be turned up when carefully following the procedures in Chapter 4.*



**Figure 2-18 Aurora Control Unit**

Power is applied to the Aurora Control Unit (ACU) when the ECU-Plus is powered up. Failure to observe all **CAUTION** notices regarding the controls on the ACU may result in damage to the PMT and probe tip.

The light controls refer to the light on the Aurora-2 head that illuminates the sample area. This light is only used in procedures that involve the video monitor and should *not* be on during NSOM scanning or any time the PMT voltage is turned up.

Note that the Feedback switch is not connected in the current configuration.

## OPTICS CONTROLS

### TRANSMISSION OBJECTIVE

The transmission objective, positioned directly beneath the sample stage, is manually adjusted in X, Y, and Z (focus) with the translation knobs shown in Figure 2-19.



Figure 2-19 Transmission Objective Translation Knobs

### FLIPPER MIRRORS

The two flipper mirrors can be flipped up or down to select the desired path of the beam leaving the reflection tower. The beam can be directed to the CCD camera, the PMT, or to optional hardware.

Coarse adjustment of each mirror is performed by loosening the screw underneath the supporting column. Use the yoke supplied with the Aurora-2 to hold the mirror temporarily in place while the position is adjusted. Once the position is set, tighten the screw under the base of the column.

The X and Y fine-adjust screws are on the mount.

### ROTATING MIRROR

The rotating mirror is controlled by a knob on the front of the stage, as shown in Figure 2-20. The knob has two positions: the counter-clockwise limit selects the reflection objective; the clockwise limit selects the transmission objective.

### REFLECTION OBJECTIVE

The reflection objective is focused by manual rotation of the lens. The manual tilt and swivel translation knobs are shown in Figure 2-20.

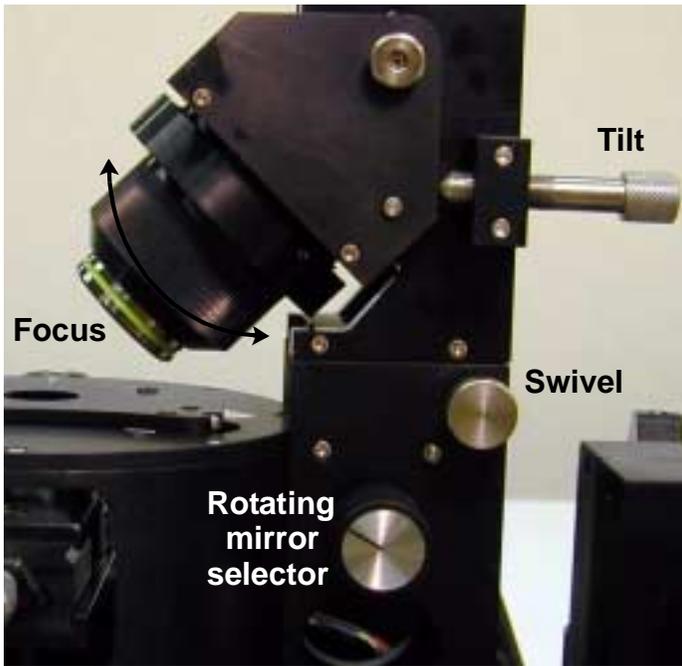


Figure 2-20 Reflection Tower Controls

## OPTICS SET-UP

### COUPLING THE LASER INTO THE FIBER

**WARNING**  *Always use caution when operating the laser. NEVER look directly into the laser aperture, fiber end, or probe tip. This can cause severe eye damage and even blindness. Follow all the safety precautions and instructions that pertain to use of the laser and laser coupler.*

Two pieces of fiber-optic cable are used in NSOM scanning: the laser fiber (the piece that runs from the laser to the cam splicer) and the tip fiber (the piece that runs from the cam splicer to the probe). The instructions in this section describe how to couple the laser into the laser fiber. These instructions apply to the laser and laser coupler sold as options by ThermoMicroscopes.

The laser coupler consists of two translation stages: one for the fiber chuck, which can be moved in the X, Y, and Z (focus) directions, and one for the fine adjustment lens located between the objective and the laser.

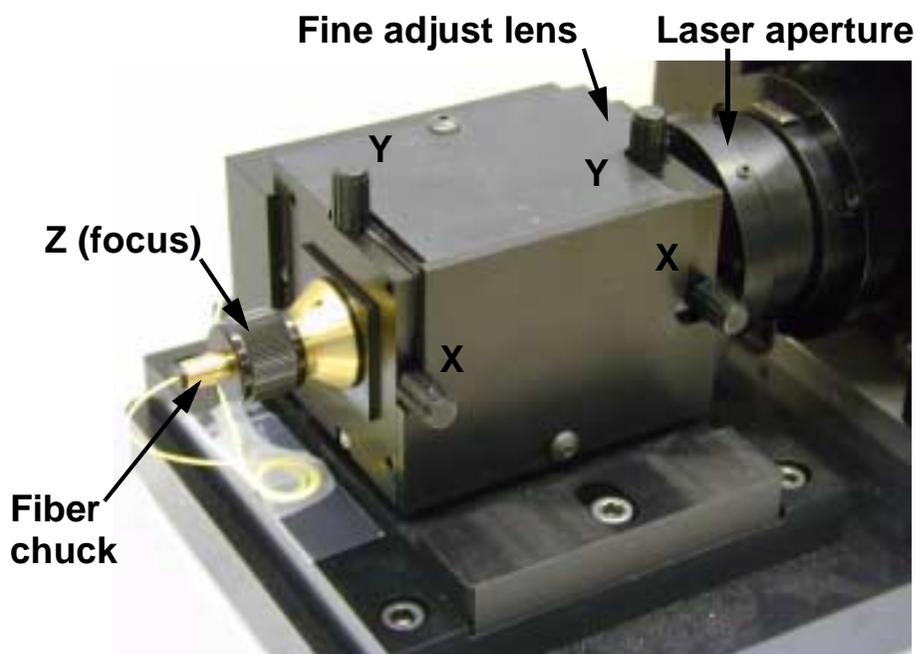


Figure 2-21 Laser Coupler

The fiber chuck is adjusted when a new fiber is inserted or when a large adjustment of the coupling is necessary. The fine adjustment (closest to the laser) is used to make a small adjustment to the coupling after it has been optimized.

The procedures for inserting a fiber into the coupler and coarse adjustment are generally performed only once, by the service engineer when the instrument is installed. However, these instructions are included so the user can adjust the coupling to achieve maximum power transfer into the fiber, or in the event that a new fiber is used. The fine adjustment of the coupling is performed more often.

To better understand the fiber coupler, you can remove the cover, as instructed in the section below, and follow along with the descriptions of the various components.

---

**WARNING**  *Make sure the laser is turned off when the cover to the laser coupler is removed, or wear the appropriate laser safety goggles. **NEVER** look directly into the fiber tip or laser aperture! Severe eye damage or blindness could result.*

---

## INSERTING A FIBER IN THE COUPLER

- Step 1** Remove the cover from the coupler. You will see that the fiber is held by a brass coupler chuck. The chuck is held in position by a chuck clamp. The clamp can be translated in the X, Y, and Z to position the fiber in the laser beam.
- Step 2** When it is time to use a new fiber, remove the coupler chuck from its clamp by unscrewing the lock collar on the end of the coupler and removing the old fiber.
- Step 3** Feed the new fiber through the chuck far enough so you can strip and cleave the end of the fiber.
- Step 4** Carefully strip about 10mm of the polymer buffer off the fiber.
- Step 5** Cleave the fiber at about 3-5mm, and pull the new cable back into the chuck, with about 5mm of fiber extending from the lock collar. (More detailed instructions on how to cleave the fiber are given on page 2-22.)
- Step 6** Replace the chuck in the chuck clamp, with the cleaved end of the fiber facing the laser objective lens. The distance between the cleaved end of the fiber and the objective lens should be approximately 3mm.
- Step 7** Tighten the chuck clamp to hold the chuck securely.
- Step 8** Replace the cover.
- Step 9** Temporarily secure the other end of the fiber in a way that will not pose a risk of laser light exposure to anyone's eyes. Tape it to the Aurora-2 base plate or gently seat it into the fiber holder on the end of the cam splicer assembly.

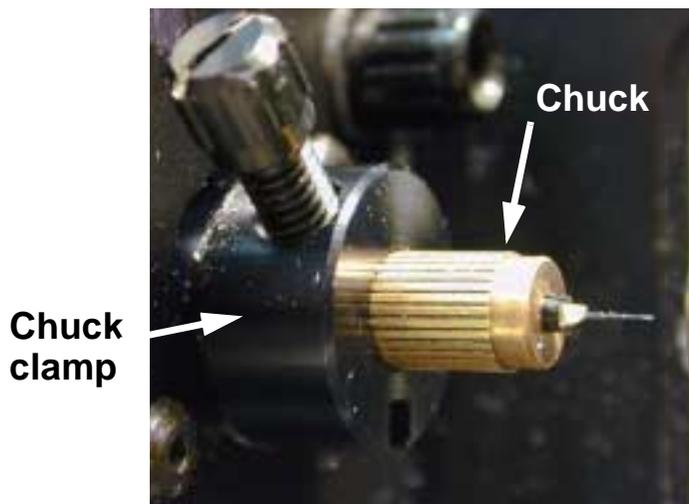
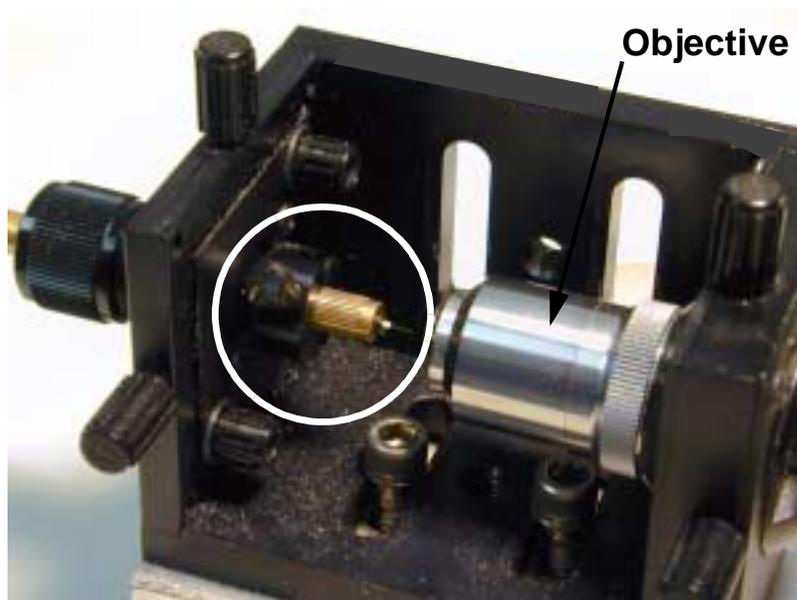


Figure 2-22 Coupler Chuck and Clamp

### COARSE ADJUSTMENT

- Step 1** Begin by turning the power adjust control on the laser to its lowest setting (to the counterclockwise limit), then turn on the power to the laser. The laser is operated on the low setting. High power settings may introduce unwanted noise.
- Step 2** The fiber end should be approximately 3 mm from the objective lens. This is the focal length for the lens. The maximum intensity will be at this 3 mm position.
- Step 3** Aim the free end of the fiber at a flat surface, such as a table, wall, or index card. Watch the intensity change as the coupler X-Y translation controls are adjusted.

**Step 4** The Z axis—the fiber focus ring—may need to be slightly adjusted for maximum power. The focus is correct when small changes in X and Y translation produce large changes in intensity. The fiber is out of the lens focal region when large X and Y translation produces little change in light intensity. After adjusting the Z axis, it is usually necessary to re-adjust the X and Y translation.

**Step 5** Adjust the X axis for maximum light intensity, then adjust the Y axis. Go back and re-check the X axis, then re-check Y. Go back to Step 4 if the Z axis needs further optimization.

## FINE ADJUSTMENT

Use the X-Y fine adjustment closest to the laser to optimize the coupling.

## COUPLING THE LASER LIGHT INTO THE PROBE

---

**WARNING**  *NEVER look directly into the fiber tip or laser aperture! Severe eye damage or blindness could result.*

---

Coupling the laser light into the probe is a procedure that is performed frequently. It can be repeated any time it seems the intensity at the tip could be improved.

The tip fiber should be stripped and temporarily installed in the cam splicer, as described on page 2-14.

---

**CAUTION**  *Do not follow this procedure when the tip is in feedback.*

---

**Step 1** Use the fiber stripper tool to strip away about 25mm of the polymer buffer off the end of the laser fiber. Note that once stripped, the fiber is very brittle.

**Step 2** Use the optical fiber cleaver from the tool kit to cleave the end of the laser fiber (you may also want to refer to the documentation supplied with the fiber cleaver):

- (a) Open the jaws of the cleaver, and seat the fiber into the notch in the lower jaw.
- (b) Insert the fiber until the stripped portion on the end aligns with the 16 mm mark.
- (c) Press the scoring bar down on the fiber. Take care not to break the fiber.
- (d) Gently flex the tongue of the cleaver down until the fiber cleaves. Take care not to touch the tip.

**Step 3** Carefully insert the laser fiber into the cam splicer at the opposite end from the tip fiber. To help the laser fiber enter, gently roll or twist it as it is inserted. Insert the laser fiber far enough to touch the tip fiber. Pushing on one fiber should move the other fiber.

**Step 4** Put the splicer arms in the locked (down) position.

**Step 5** Gently seat the laser fiber into the fiber holder on the end of the cam splicer assembly.

**Step 6** Check to see that laser light is coupled into the tip fibertip fiber. The laser light may not be visible at the tip, but if the room lights are turned off, the tip fiber should be glowing. If there is no light visible in the tip fiber, repeat Steps 2-5.

Note that once the tip has been coupled to the laser, thermal expansion of the tip will take place for the next few minutes. It is a good idea to wait approximately 15 to 20 minutes before attempting a scan, to allow the tip to reach thermal equilibrium.

## OPTICAL TRAIN ALIGNMENT

The Aurora-2's optical train has been adjusted at the factory, and the service representative who installed the system has made final adjustment corrections. However, the user should be able to make adjustments to the optical path if necessary.

The objective of this procedure is to have the center of the optical beam pass through the center of each optical element in its path. This means that the image beam should be centered on both the transmission and reflection lenses, the beam mirrors, the prisms, the focusing lenses, and the CCD camera and PMT detector faces (see Figure 2-2 on page 2). This is essentially the same goal as in any optical system, and the procedures for achieving this goal are similar.

Precise optical adjustment is critical for NSOM imaging and any advanced techniques. Because the tip emits little light, the optical path must be accurately focused to collect the maximum amount of light with the detector. If the detector signal strength is very low, make sure that none of the beam is being lost along the path and that the beam is focused on the detector.

For this procedure, you will need a small piece of paper to use as a "beam finder." The back of a business card or index card works well for this.

---

**CAUTION**  *Keep the PMT Voltage on the Aurora Control Unit set to 0, unless the PMT is shielded from all sources of light (except the laser during NSOM imaging). Exposure to light can damage the PMT. The voltage should only be turned up when carefully following the procedures in Chapter 4.*

---

- Step 1** Set up the Aurora-2 as described in the above procedures in this chapter. It is assumed that the user is familiar with removing and replacing the head and mounting a sample.
- Step 2** If the system is not powered up, first turn on the ECU-Plus, then the computer and the video monitor. (The ECU-Plus should always be turned on before the computer so that the ECU interface is recognized and initialized.)
- Step 3** Open the SPMLab software, and enter the Data Acquisition module.
- Step 4** Turn on the light on the Aurora-2 head with the switch on the Aurora Control Unit. This light illuminates the sample during tip positioning and focusing.
- Step 5** Remove the head from the stage, if necessary.
- Step 6** Mount the ThermoMicroscopes NSOM standard sample on the stage. The easiest way to do this is by using a few small pieces of tape at the corners of the sample. Or, you can put a small drop of immersion oil underneath the sample.
- Step 7** Turn the Z-height thumbscrews on the head one full turn clockwise, and carefully place the Aurora-2 head on the stage, lowering the head slowly to make sure the tip does not touch the sample when it comes to rest in the kinematic mount. The tip should now appear in the video monitor.

The tip may be difficult to find in the reflection optics if it is too far from the sample. Check the tip-sample area by eye to get an idea of the distance, and use the manual thumbscrews to lower the head, if necessary, being careful not to crash the tip into the sample.

- Step 8** Use the video monitor to focus the image in both transmission and reflection modes. Use the rotating mirror to switch back and forth between these images. The tip should appear in the middle of the monitor. Use the appropriate focus controls for the transmission and reflection objectives.
- Step 9** Rotate the mirror to reflection mode, and focus the optics on the sample. This image should be the more easily recognizable of the two.
- Step 10** Click on the Tip Down button  on the Data Acquisition tool bar to lower the tip until it is within a few millimeters of the stage, as seen on the video monitor.
- Step 11** Focus the reflection optics on the tip.
- Step 12** Rotate the mirror to transmission mode, and focus the optics on the sample.
- Step 13** Once both transmission and reflection mode images are focused, look at the image on the video monitor. The image should fill the monitor, and no edges should be visible. If the edges can be seen, use the X and Y fine adjust screws on the camera path flipper mirror mount to center the image. Be sure to check both transmission and reflection images after making an adjustment.
- Step 14** Flip the mirror to the down position when finished centering the image.
- Step 15** Using the “beam finder,” find the beam leaving the reflection tower. Make sure the beam is hitting the PMT path mirror. This mirror’s coarse position can be adjusted by loosening the screw underneath the supporting column. Use the yoke supplied with the Aurora-2 to hold the mirror temporarily in place while the position is adjusted. Once the position is set, tighten the screw under the base of the column.
- Step 16** Trace the beam from the path mirror to the focusing lens. Make sure the beam intersects the middle of the lens. If needed, the lens position can be adjusted the same way the mirror’s position was.
- Step 17** Make sure the beam is focused on the center of the PMT’s detector window. Slide the lens back and forth so the beam is focused as nearly as possible on the tube aperture. Or, you can unscrew the PMT from its support bracket (4 screws).

Once adjusted, the instrument should not require major optical path alignment. The instrument is now ready to take a scan.

---

## Chapter 3

# Taking a Topographic Image

### **PROCEDURE OVERVIEW**

NSOM combines both topography and optical imaging. It is common to take a topographic image first to allow the user to set up the scan parameters and potentially identify features of interest on the sample. Then the laser is coupled into the tip, and a combined topographic and optical scan is taken.

Before taking an NSOM image, it is essential to master taking a topographic image, which is nearly identical to non-contact Atomic Force Microscopy (AFM) techniques. Topographic scanning requires learning the techniques for bringing the tip into feedback. When the tip is in feedback, it can track the topography of the sample, thus it is in the near-field range of the sample, which is necessary for NSOM imaging.

Chapters 3 and 4 cover this two-part technique for taking a scan of the standard sample included with the instrument. The combined scan techniques described below will be applicable to any sample.

The instructions in this chapter assume that the Aurora-2 has already been set-up as described in Chapter 2. All components should be powered up, and the Data Acquisition module of the SPMLab software should be open.

## APPROACHING THE SAMPLE

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**CAUTION**  *Be careful when lowering the tip not to crash it into the sample surface.*

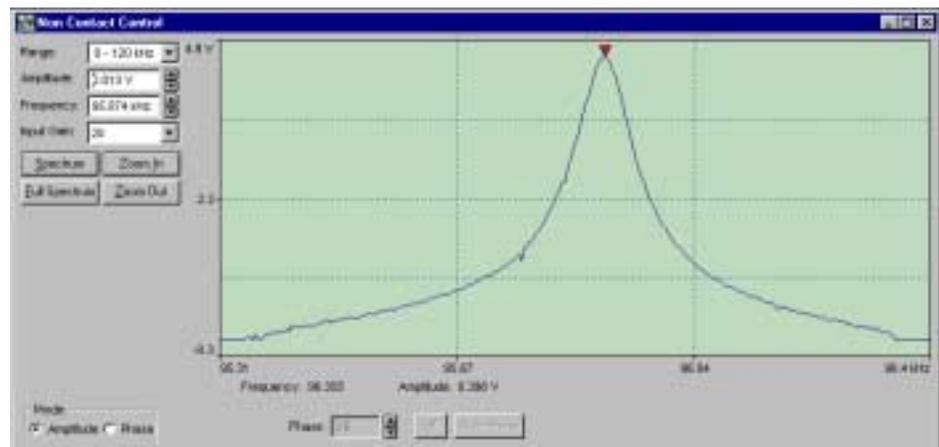
Feedback refers to the attenuation of the tip's oscillation amplitude. When the tip is in feedback at the sample surface, the sample topography can be tracked and a topographic image taken. This also means that the tip is in the near-field range, enabling NSOM imaging. A technique known as "false feedback" will be used to bring the tip into the near field. This allows for a gentle yet efficient approach, whereas using the Z-motor to approach the sample runs the risk of breaking the tip and is usually very slow.

### FINDING THE RESONANT DRIVE FREQUENCY

For each probe, it is necessary to set the drive frequency at the resonant frequency of the tuning fork. The objective is to maximize the magnitude of the tuning fork vibration by adjusting the driving frequency control.

For the following procedure, refer to the "Non Contact Operation" section in Chapter 2 of the *SPMLab Software Reference Manual*. When the SPMLab software is configured for the Aurora-2, the Non Contact window is automatically open, with the Active mode checked, upon entering the Data Acquisition mode.

- Step 1** Select "Internal Sensor, Feedback" from the drop-down signal source list to the right of the upper oscilloscope.
- Step 2** Select **Amplitude** from the Mode group box in the Non Contact window.
- Step 3** In the Non Contact window, enter the following starting parameters:
  - Amplitude = .1 V (drive amplitude)
  - Input Gain = 8. (input gain of the I/O MOD+ board)
- Step 4** In the Signal window, select **SCOPE** mode.
- Step 5** From the Range drop-down list, select the frequency range that is closest to 90-100 kHz, which is typical for the tuning fork.
- Step 6** In the Non Contact window, select **SPECTRUM** to run a frequency sweep.
- Step 7** Zoom in on the peak so that the frequency window is 2 kHz (or less) wide.
- Step 8** Make sure the peak is not cut off. The maximum voltage range of this window is 10 V. If the peak is cut off, reduce the Amplitude. Ideally, the peak will sit at around 5 V, which corresponds to about -30nA on the internal sensor signal. Fine tuning will be done later.
- Step 9** Click on the peak to select the drive frequency, which will then appear in the **DRIVE FREQUENCY** field. See Figure 3-1.



**Figure 3-1** Selecting the Drive Frequency

- Step 10** Select **PHASE** from the Mode group box in the Non Contact window, and watch the internal sensor signal.
- Step 11** Click on the **AUTO PHASE** button in the Non Contact window. The internal sensor signal should go to 0.
- Step 12** Click on the **+90** button three times (for a total of 270 degrees) to find the phase that shows the most negative value for the internal sensor signal.
- Step 13** Fine tune the Non Contact controls using the following guidelines:

The aim is to have the drive amplitude be as low as possible while keeping the internal sensor signal at approximately the same value and the RMS noise (displayed in blue in the Signal window) less than 0.1.

Reducing the drive amplitude will cause the internal sensor signal to increase (move closer to 0). Increasing the gain will bring the internal sensor signal down (more negative, i.e., further from 0). However, increasing the gain will proportionally increase the RMS noise.

- Step 14** Once the Non Contact controls have been fine-tuned, minimize the Non Contact window. To adjust these settings, the tip must be out of feedback. These settings will be saved when you exit the SPMLab software.

## MOVING THE TIP INTO FEEDBACK

- Step 1** Set the rotating mirror to reflection mode.
- Step 2** Use the thumbscrews to lower the head. Bring the tip within a few millimeters of the surface.
- Step 3** Lower the set point below the current sensor signal level. For example, if the sensor signal level is -30 nA, lower the setpoint to -50 or -60 nA.
- Step 4** Select “Z-Piezo” from the drop down signal list to the right of the lower oscilloscope to monitor the z-piezo voltage.

---

**CAUTION** ⚠ *Do not leave the z-piezo in its fully retracted position (-220 V), as this causes it to wear out very quickly.*

---

**Step 5** Click the **TIP APPROACH** button on the Acquisition Control Panel. The z-piezo should fully retract, as shown by a z-piezo voltage of -220 V. The tip is now in “false feedback.”

**Step 6** Watch the video monitor while lowering the tip with one of the two manual thumbscrews. Bring the tip as close to the sample surface as possible without crashing the tip.

It may be useful to monitor the tip both with the light on and off. When the light is off, and the laser is coupled into the probe fiber, the tip appears as a bright spot and will be reflected on the sample’s surface. When the light is on, a dark image of the tip and the length of the fiber is visible, and the image will be reflected on the sample’s surface.

**Step 7** Set the P-I-D settings to:

$$P \approx 1$$

$$I \approx 0.04$$

$$D = 0$$

The P-I-D settings should be low (the Integral gain in particular) so the tip doesn’t jump as it approaches the sample.

**Step 8** Click on the **SET** button to the right of the internal sensor signal. The current internal sensor signal value (on the horizontal line at the middle of the window) can now be read clearly.

**Step 9** Enter the internal sensor signal value (or a slightly lower value, to be safe) in the **SET POINT** field.

**Step 10** Click the up arrow next to the **SET POINT** field to raise the value in small increments until the z-piezo moves.

---

**CAUTION** ⚠ *Never turn the light on or off when the tip is in feedback at the surface. A voltage spike could cause a tip break.*

---

---

**CAUTION** ⚠ *Do not power down the electronics while the tip is in feedback at the surface.*

---

**Step 11** Monitor the z-piezo voltage. If the tip was within scanner range of the surface, the z-piezo voltage should steadily rise and stabilize at some arbitrary value, somewhere between -220 and 220 V, dependant on how far the tip was from the sample surface. The tip is now in feedback at the surface (“true feedback”), and you can skip to the next section to take a topography scan.

- Step 12** If the z-piezo signal reaches full deflection, 220 V, the tip was not within scanner range of the surface. Retract the scanner with the z-piezo by setting the setpoint significantly below the sensor signal value (i.e., -50 or -60 nA). Watch the z-piezo signal voltage, and make sure it drops to -220 V.
- Step 13** Watch the video monitor image while using the thumbscrews or the z motor (Tip Down button ) to carefully lower the tip. Be very careful not to crash the tip into the surface.
- Step 14** Enter the internal sensor signal value (or a slightly lower value, to be safe) in the **SET POINT** field.
- Step 15** Click the up arrow next to the **SET POINT** field to raise the setpoint above the current sensor signal value again. Monitor the z-piezo voltage as the tip nears the surface. Again, the z-piezo voltage should stabilize somewhere between -220 and 220 V.
- Step 16** If the piezo reaches full deflection, repeat Steps 12-15 until the z-piezo voltage stabilizes within the voltage range.

## TAKING A TOPOGRAPHY SCAN

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**CAUTION**  *To avoid crashing the tip and damaging both the tip and sample, the P-I-D settings must be set properly.*

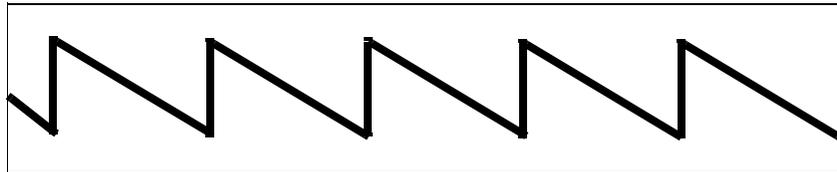
Once the tip is in feedback with the surface, the P-I-D settings are adjusted to optimize the scan. These settings tune the distance control feedback circuit to respond quickly and accurately to changes in surface topography. This gives the best topographic imaging.

### OPTIMIZING THE P-I-D SETTINGS

- Step 1** Set the Integral gain between 0.08 and 0.30. This range might change depending on the sample and the specific tip.
- Step 2** Click the **LINE** scan option in the upper right of the Signal window. This scan mode lets the user evaluate the changes to the internal sensor and topography signals as the scan rate, setpoint, and P-I-D settings are adjusted. Line scan mode scans over the same line on the sample and displays three overlaid profiles, each displayed in a different color. This display allows the user to adjust the scan parameters to get a stable, repeatable scan.
- Step 3** Select “Internal Sensor, Feedback” (upper oscilloscope) and “Topography” (lower oscilloscope) from the drop-down signal source lists in the Signal window.
- Step 4** Monitor the signal while you perform a line scan. If the Integral gain setting is too high, you will notice oscillations in one or both signals.
- Step 5** Set the Proportional gain to 1. If the system oscillates, lower the Proportional gain until the oscillation stops, then raise it slowly. Usually the Proportional gain is set between 0.5 and 1.
- Step 6** Make sure the Derivative gain is set to 0 (this setting is generally not used).

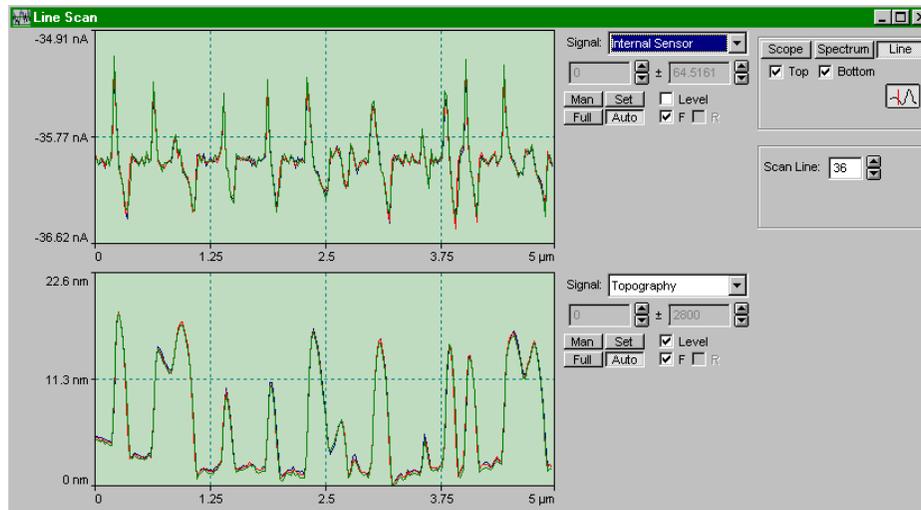
**Step 7** Set the scan rate to about 1/2 the scan range, e.g., if the range is set to 20  $\mu\text{m}$ , the scan rate would be about 10  $\mu\text{m}/\text{s}$ . The scan rate can be a very important parameter in obtaining a stable signal. If the sample has high topographic features, and a stable signal cannot be obtained by lowering the P-I-D settings, reduce the scan rate.

If the line scan has a “sawtooth” shape, as shown in Figure 3-2, with the left slope very steep and the right slope relatively flat, it is typically an indication that the tip is too far from the surface, or that the P-I-D settings are too low.



**Figure 3-2** “Sawtooth” Line Scan Shape

**Step 8** When the P-I-D settings and set point are set properly, you will see a quiet, stable signal, as shown in Figure 3-3.



**Figure 3-3** Properly Adjusted P-I-D Settings and Set Point

When a scan begins, the line scan of the internal sensor signal will no longer be displayed. To view the internal sensor signal, set the Signal window to **SCOPE** mode, and open an internal sensor signal window: select **SETUP**  $\Rightarrow$  **ACQUIRE**, and from the Data Channels list, check the Fwd checkbox for the Internal Sensor channel (this can not be done when scanning is in progress).

## ENDING A TOPOGRAPHY SESSION

**Step 9** Take a scan of the sample by clicking the Instant Scan button  on the Data Acquisition tool bar. Adjustments to the P-I-D settings can be made while the scan is being taken. There is no need to restart the scan.

To take an NSOM scan, leave the tip in feedback and proceed to Chapter 4, as all the signal parameters are already adjusted correctly. Note that it is important to attain some proficiency in taking topographic scans before taking an NSOM scan.

---

**CAUTION**  *Do not power down the electronics while the tip is in feedback at the surface.*

---

**Step 1** Click the Instant Scan button  to stop scanning.

**Step 2** Click on the Tip Up  button on the Data Acquisition tool bar to raise the tip away from the sample, bringing the tip out of feedback.

**Step 3** Exit the SPMLab software.



---

## Chapter 4

# Taking an NSOM Scan

Before following this NSOM scanning procedure, the Aurora-2 should be set up as described in Chapter 2, and you should have taken a topography scan following the procedure in Chapter 3. In order to take an NSOM scan, it is important to attain some proficiency in taking topographic scans.

---

**CAUTION**  *The Photomultiplier Tube (PMT) can be damaged by repeated exposure to high light levels with the voltage supplied to the tube. More importantly, repeated exposure will degrade the PMT's sensitivity over time, resulting in degraded performance. It is a good practice to protect the PMT from high light levels, which helps ensure long detector lifetime. When concluding an NSOM session, wait 10-15 seconds before exposing the PMT to light. In general, whenever the voltage is supplied to the tube, the PMT should be in a dark environment.*

---

## CHECKING FEEDBACK PARAMETERS

---

These parameters should still be set correctly from the topographic scan. However, as the tip heats and expands, some adjustments might have to be made to the P-I-D settings. Also, it is a good idea to recheck the tuning fork resonance frequency. Following the guidelines in Chapter 3, adjust the P-I-D settings for the best signal. Recheck these settings immediately before beginning the scan.

## APPROACHING THE SAMPLE AND TAKING A SCAN

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Refer to Chapter 3, observing all Warning and Caution statements, when following this procedure.

- Step 1** Locate the tip with the transmission and reflection objectives.
- Step 2** Bring the tip into feedback, if it is not already, following the procedure under “*Moving the Tip into Feedback*,” on pg. 3-3.
- Step 3** The z piezo voltage should be around 0.
- Step 4** Pull the z piezo back by typing in a set-point value that is more negative than the internal sensor signal. E.g., if the internal sensor signal is -30 nA, type in -64 nA. The piezo will retract. This is done so the tip doesn’t break when you flip the mirror.
- Step 5** Make any final necessary adjustments to the transmission objective. The image should be in focus and in the middle of the monitor.
- Step 6** Flip the camera path mirror down, to open the PMT path.
- Step 7** Close the instrument enclosure and, if possible, dim or turn off the room lights.
- Step 8** Turn off the Aurora-2 light.

---

**CAUTION**  *The light on the Aurora-2 must be off to prevent damage to the PMT.*

---

- Step 9** Select **SETUP** ⇒ **ACQUIRE**, and from the Data Channels list check the **FWD** checkbox for the NSOM channel.
- Step 10** Bring the tip back into “true feedback,” i.e., at the sample surface, as described in Steps 7-16 under “*Moving the Tip into Feedback*,” on pg. 3-3. The P-I-D settings should be low (the Integral gain in particular) so the tip doesn’t jump as it approaches the sample. Be sure to observe *all* the Warning and Caution statements!
- Step 11** Increase the P-I-D settings to improve the topography response, following the procedure under “*Optimizing the P-I-D Settings*,” on pg. 3-5.
- Step 12** Click the **SCOPE** option in the upper right of the Signal window, and choose “NSOM” from the drop-down signal source list to the right of the upper oscilloscope. The signal should read 0, both on the oscilloscope and the Aurora Control Unit.
- Step 13** Slowly turn up the PMT voltage on the Aurora Control Unit. The PMT will typically “turn on” at around 230 V. Watch the PMT signal, and make sure there is a signal response as the PMT voltage is increased. Aim for a signal of about 200mV. Increasing voltage increases the signal strength, but it also increases the noise level within the signal.
- Step 14** Perform a line scan by clicking the **LINE** option in the Signal window.
- Step 15** Adjust the set point and the P-I-D settings until you get repeatable line scans, as shown in Figure 4-1.

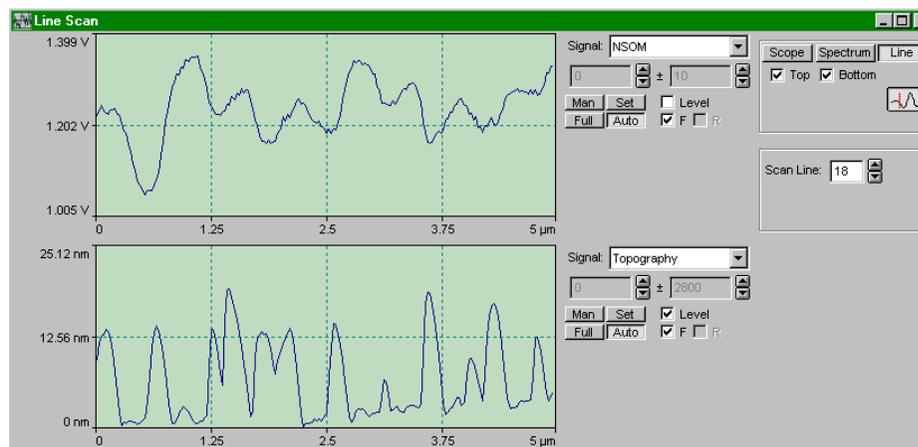


Figure 4-1 Optimized Line Scans

**CAUTION** ⚠ Lower the PMT voltage immediately if you notice a sudden, dramatic increase of the NSOM signal (indicating a tip break). Exposure to these high light levels will permanently damage the PMT.

**Step 16** Begin a scan by clicking the Instant Scan button  on the Data Acquisition tool bar. Adjustments to the P-I-D settings and set point can be made while the scan is being taken. There is no need to restart the scan.

During the scan, you can keep the oscilloscope in **LINE** mode and make slight corrections as needed to compensate for any erratic phenomena, e.g., drift, indicated by a “sawtooth” pattern on the Topography trace (left slope much steeper than the right slope). Or, it is possible for the tip to pick up a loose particle from the sample surface. Such events are not unusual, and there is no need to stop the scan and start over; simply make corrections.

## ENDING AN NSOM SESSION

**CAUTION** ⚠ When concluding an NSOM session, wait 10-15 seconds before exposing the PMT to light. In general, whenever the voltage is supplied to the tube, the PMT should be in a dark environment.

- Step 1** Click the Instant Scan button  to stop scanning.
- Step 2** Click on the Tip Up button  on the Data Acquisition tool bar to raise the tip away from the sample, bringing the tip out of feedback.
- Step 3** Turn the PMT voltage all the way down to 0.

**Step 4** *Wait 10-15 seconds* before exposing the PMT to any light.

**Step 5** Exit the SPMLab software.

The scanned images can be saved and processed using the SPMLab software, which contains a number of analysis and image processing tools. Refer to the *SPMLab Software Reference Manual* for further details.

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## Chapter 5

# Counter Board Operation

### INTRODUCTION

This chapter describes how to operate the Aurora-2 with the optional I/O-P photon counter board. The counter board is used in combination with a photon-counting detector (purchased from a third-party manufacturer), either a PMT or an avalanche photo diode (APD) module. The user should be familiar with the general operation of the Aurora-2 before attempting photon counting operation.

### HARDWARE SET-UP

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**CAUTION**  *Make sure the power is OFF to all the modules and the computer while setting up. Connecting cables to powered-up electronics may damage the modules.*

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**CAUTION**  *Do not activate your light detector when it is exposed to room light, as this can cause damage.*

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- Step 1** Connect a BNC cable from the output of your detector to Input 1 or Input 2 of the I/O-P.
- Step 2** Make the necessary power connection to your detector in accordance with the user manual.
- Step 3** Make sure that all cable connections are properly secured.
- Step 4** Optimize the alignment of the detector. See “*Optical Train Alignment*,” in Chapter 2 for guidelines on alignment.

## COUNTER SET-UP

- Step 1** Once all the connections have been made, power up your system (first turn on the ECU-Plus, then the computer and the video monitor).
- Step 2** In order to activate the counter software, the **[Aurora]** section of the **stages.ini** file must include the following entries:
- ```
CounterBoard=Yes
Line5=Counter_1, InActive, Counter1, NonInverted
Line6=Counter_2, InActive, Counter2, NonInverted
```
- Step 3** Open the SPMLab software, and enter the Data Acquisition module.
- Step 4** Select **SETUP ⇒ COUNTER**. The Counter Setup dialog box opens, as shown in Figure 5-1.

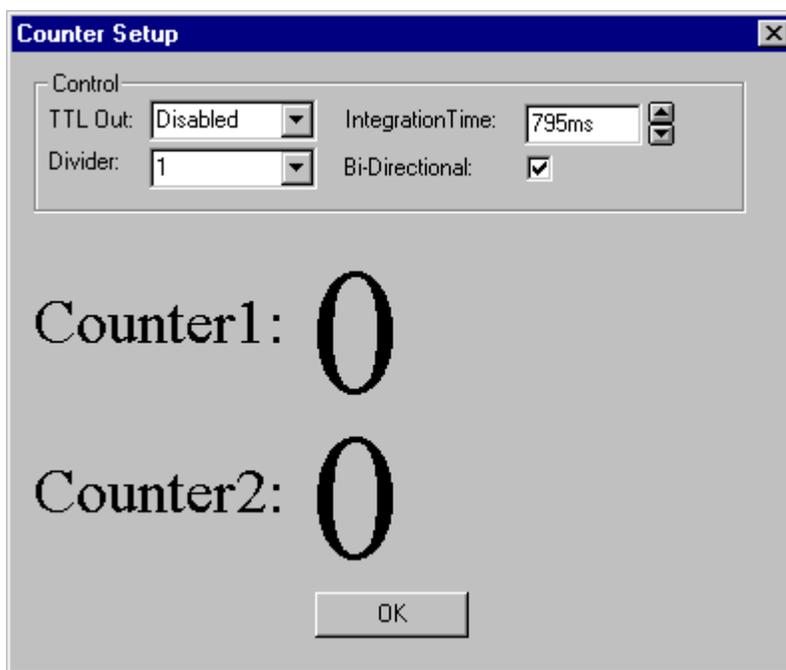


Figure 5-1 Counter Setup Dialog Box

**TTL OUT** This setting determines where the ECU-Plus outputs a synchronization pulse.

**DISABLED** Counting is not active, and both counters will display zero. Use this setting for running the non-photon-counting PMT or for any other scanning that does not require the counter. When the counter board is disabled, the scanning routine will disregard the integration time setting.

**OUT1 (BNC1)** Activates the counter board so the software can process the counts. When the counters are active, the scanning routine changes: the tip stays at each pixel for the selected integration time.

**OUT2 (BNC2)** This setting is currently disabled.

**DIGI** This setting is currently disabled.

**NONE** The counter board is enabled, but there is no TTL output.

## INTEGRATION TIME

This number determines the counting interval in milliseconds. During scanning, the software will adapt in accordance with the selected integration time, i.e., the tip will remain at each pixel for the time selected, regardless of the selected scan speed. Therefore, it is not necessary to synchronize the scan.

## BI-DIRECTIONAL

When checked, the counting routine will work for the forward and reverse scan. When unchecked, the counting will occur only during the forward scan, and the selected scan rate value will be used as the scan speed for the reverse direction.

## DIVIDER

The number of counts can be divided by up to 8 bits.

## OPERATIONAL CONSIDERATIONS

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**CAUTION**  *To prevent damage to your APD module or photon-counting PMT, do not exceed the maximum count rate. Refer to the manufacturer specification.*

You can open a scanning window for each counter to display the counts. When the counters are active, **Counter\_1** and **Counter\_2** will be available as Data Channels when you select **SETUP** ⇒ **ACQUIRE** (when the counters are disabled, these choices will be grayed out). The images from these signals are saved with the extensions **xfr** and **yfr**, respectively, and can be processed like any other image file.

If the counter board is present in the system, but no signal is fed into the BNC's, both channels will read 0 counts. If one or both counters reads 65535 with no signal connected, the board is not properly installed.

The integration time can be varied from 0.1 ms to 1000 ms. Select an integration time that suits your experiment. Remember that the tip will stay at each pixel for the integration time that you selected, so scans can become slow.

If data is collected in the forward direction only, it is advisable to disable the **BI-DIRECTIONAL** checkbox to speed up scanning.

The counter uses 16 bit for displaying the data and 8 bit for scaling. If your signal exceeds 64k counts, a divider greater than 1 may be used to scale the signal. Alternatively, the integration time may be reduced to speed up scanning.



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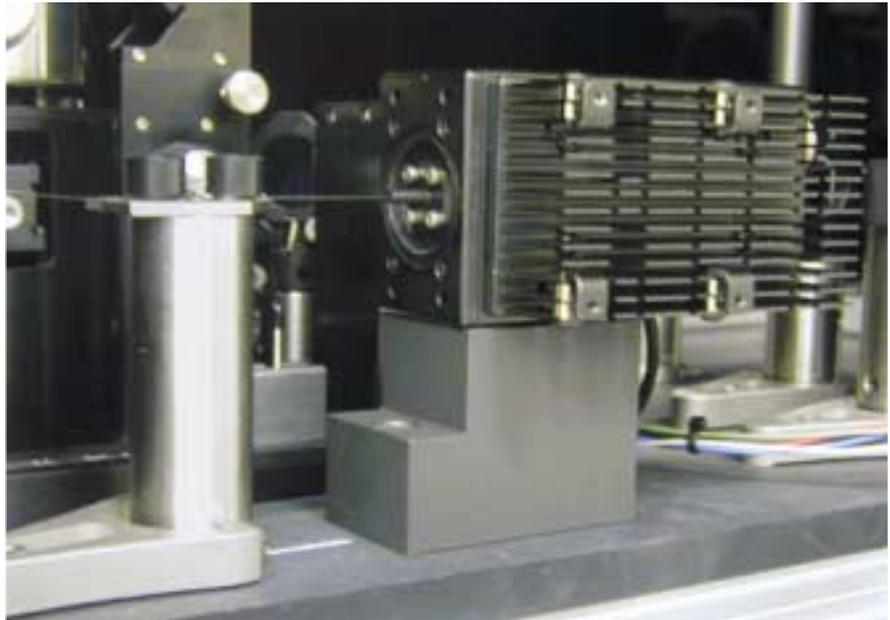
## Appendix: Collection Mode

In collection mode, the NSOM tip will be used to gather light rather than act as an illumination source. Therefore, the light exiting the tip fiber must be guided onto a suitable detector. If you are using a detector with a rather large area, such as a PMT, that might simply mean pointing the cleaved end of the fiber at the active area. However, if you are using an APD, for example, some imaging system has to be set up, as the beam exiting the fiber (NA 0.1) will simply overfill the active detector area.

The following example describes a collection mode set-up using a Hamamatsu photon-counting PMT module, as shown below.

After cleaving the fiber at approximately 3mm, put the fiber into the guiding groove of the holder and insert it carefully into the 12mm-long tube in front of the detector. Make sure not to break the fiber by touching the detector window. Secure the fiber in the guiding groove using the two magnets.

You are now ready to go into feedback and conduct your experiments.



**Collection Mode Set-up with Hamamatsu Photon Counting PMT**

